Assessing habitat suitability and connectivity for an endangered salamander complex

A thesis submitted to the Committee of Graduate Studies in Partial Fulfillment of the Requirements for the Degree of Master of Science

in the Faculty of Arts and Science

TRENT UNIVERSITY

Peterborough, Ontario, Canada

© Copyright by Graeme Smith 2022

Environmental and Life Sciences M.Sc. Graduate Program

September 2022

Abstract

Assessing habitat suitability and connectivity for an endangered salamander complex Graeme Smith

Habitat loss and fragmentation have significantly contributed to amphibian population declines, globally. Evaluating the state of remaining habitat patches can prove to be beneficial in identifying areas to prioritize in conservation efforts. Pelee Island, Ontario is home to a complex of salamanders including small-mouthed salamanders (*Ambystoma texanum*), blue-spotted salamanders (*A. laterale*) and unisexual *Ambystoma* (small-mouthed salamander dependent population). These populations have declined from intense landscape changes since the late 1800s, particularly from the historical drainage of wetlands. In this thesis, I evaluated the suitability and connectivity of habitat patches occupied by these salamanders to assess the size of, and dispersal capabilities between, remaining habitat patches. I found that there was a low amount of suitable terrestrial habitat available for this complex of salamanders, and existing habitat patches were small and isolated. Forested areas and non-breeding wetlands were considered to be suitable habitat when adjacent to existing breeding locations, suggesting that these habitats should be a focus for conservation efforts. Notably, intervention may be necessary to maintain this amphibian complex as many assemblages are isolated from one another and potential corridors currently consist of primarily unsuitable habitat. Given that much of the salamander complex is reliant on one species for reproduction, the long-term viability of this population of *Ambystoma* salamanders may rely on the enhancement of suitable habitat near current breeding sites by conservation organizations

ii

and local stakeholders. Ultimately, the approach used in this thesis emphasizes the value of evaluating habitat within a fragmented landscape to focus conservation efforts on imperilled species.

Keywords: Amphibians, landscape fragmentation, habitat suitability, connectivity, landscape resistance, unisexual

Acknowledgements

This thesis would not have been possible without the support of many people who helped me throughout my studies. I would first like to thank my supervisor Dr. Dennis Murray for giving me the opportunity to pursue this research project, and for your invaluable expertise in helping me to develop my research skills and to grow as a scientist. I am grateful for your patience and encouragement throughout the many versions of this project. I would also like to thank Dr. Thomas Hossie, for your wealth of knowledge about this complex of salamanders and Pelee Island, and for your insightful feedback. I truly appreciate you reminding me of the importance of this work when I was bogged down in data analysis. Additionally, I would like to thank Dr. Erica Nol for serving on my committee, and for your kindness and guidance.

All of the members of the Murray Lab were integral to the completion of this thesis, particularly Evan Bare, Anne-Christine Auge, Tucker Cambridge and Drs. Jenilee Gobin and Angela Eads who were essential in helping my personal growth and provided insightful advice and support. I would also like to extend my thanks to Meghan Ward and the personnel from Scales Nature Park (Bianca, Jordan, Jose, Maria, Nathan and Wayne) for their assistance in the field. There's no one I'd rather walk through dogwood thickets and poison ivy with; thank you for your patience and encouragement. Thanks also to Jill Crosthwaite from Nature Conservancy Canada and Joe Crowley from the Ministry of the Environment, Conservation and Parks for your advice and knowledge throughout this project.

I cannot begin to express my gratitude to my friends and family, who probably thought this would never end and must be glad to be reading this. I am grateful for my family's support, especially my parents for attempting to understand when I tried to explain these complicated salamanders, for your unwavering encouragement over the past several years and for inspiring my love of nature. My success would never have been possible without the support of my friends Rachal Bolger, Nicole Winckel and Ra'anaa Brown; thank you for motivating me, being willing to change our plans when my schedule got too busy, reminding me to take breaks, and for (attempting) to keep me sane. Thank you to everyone along this journey who made time to support and encourage me.

Table of Contents

List of Figures

Figure 1: Relationship between distance to breeding location (m) and predicted habitat suitability for *Ambystoma* salamanders on Pelee Island, ON, Canada (based on response curves from Maxent models). A) The relationship between distance to breeding site and predicted habitat suitability based on spatially filtered location data $(n = 476)$ for all adults and juvenile salamanders collected from 2015-2019. B) The relationship between distance to breeding site and predicted habitat suitability for three distinct *Ambystoma* genomotypes, from data collected from 2015-2018. Dotted line depicts the relationship for diploid (LT) unisexual *Ambystoma* salamanders, the dashed line depicts the relationship for triploid (LTT) unisexual *Ambystoma* salamanders, and the solid line depicts the relationship for *A. texanum* (TT) salamanders. Higher values on y-axis represent higher probability of presence.

Figure 2: A) Predicted habitat suitability for *Ambystoma* salamanders on Pelee Island, ON, Canada using spatially filtered location data $(n = 476)$ collected from 2015-2019 (as modelled in Maxent). Environmental predictor variables include distance to breeding site, land cover type, soil type, and elevation. Green indicates areas with predicted high habitat suitability and red indicates areas with low habitat suitability. B) Predicted habitat connectivity for *Ambystoma* salamanders on Pelee Island, ON, Canada produced using spatially filtered localities $(n = 476)$ and a land cover raster grid using resistance values based on Compton et al. 2007 (see Table S1). Green indicates well-connected areas and red indicate areas with low connectivity. Lines depict predicted least-cost paths among known communities, with paths under 1 km in length depicted in purple, and paths are over 1 km in black.

Figure 3: Predicted habitat suitability for three *Ambsytoma* salamander genomotypes on Pelee Island, ON, Canada, using spatially filtered location data (from 2015-2018) and environmental predictor variables including distance to breeding site, land cover type, soil type, and elevation. Left panel: Predicted habitat suitability for *Ambystoma texanum* (TT) salamanders. Middle panel: Predicted habitat suitability for diploid (LT) unisexual *Ambystoma* salamanders. Right panel: Predicted habitat suitability for triploid (LTT) unisexual *Ambystoma* salamanders. Green indicates areas with predicted high habitat suitability, with orange and red indicating areas with low habitat suitability.

Figure 4: Predicted habitat connectivity for three *Ambystoma* salamander genomotypes on Pelee Island, ON, Canada. The models used spatially filtered localities (from 2015-2018) and a land cover raster grid with resistance values based on Compton et al. 2007 (see Table S1). Left panel: Connectivity for *A. texanum* (TT) salamanders. Middle panel: Connectivity for diploid (LT) unisexual *Ambystoma* salamanders. Right panel: Connectivity for triploid (LTT) unisexual *Ambystoma* salamanders. Green indicates well-connected areas, with orange and red indicating areas with low connectivity.

Figure S1: Predicted habitat suitability for adult and juvenile salamanders on Pelee Island, ON, Canada, using spatially filtered localities (from 2015-2019) and environmental predictor variables, including distance to breeding site, land cover type, soil type, and elevation. Left panel: Predicted habitat suitability based on all adult *Ambystoma* salamander locations (*n* = 369). Right panel: Predicted habitat suitability based on all juvenile *Ambystoma* salamander locations (*n* = 179). Green indicates areas with predicted high habitat suitability and red indicate areas with low habitat suitability.

Figure S2: Predicted habitat suitability for three seasons on Pelee Island, ON, Canada, using spatially filtered localities (from 2015-2019) and environmental predictor variables, including distance to breeding site, land cover type, soil type, and elevation. Left panel: Predicted habitat suitability based on *Ambystoma* salamander locations from March and April 2015-2019 (*n* = 190). Center panel: Predicted habitat suitability based on *Ambystoma* salamander locations from May – July 2015-2019 (*n* = 233). Right panel: Predicted habitat suitability based on *Ambystoma* salamander locations from August and September 2015-2019 (*n* = 136). Green indicates areas with predicted high habitat suitability and red indicates areas with low habitat suitability.

Figure S3: Predicted habitat connectivity for adult and juvenile life stages on Pelee Island, ON, Canada, using spatially filtered localities (from 2015-2019) and a land cover raster grid using resistance values based on Compton et al. 2007 (see Table S1). Left panel: Connectivity based on adult *Ambystoma* salamander locations (*n* = 369). Right panel: Connectivity based on juvenile *Ambystoma* salamander locations (*n* = 179). Green indicates well-connected areas and red indicates areas with low connectivity.

Figure S4: Predicted habitat connectivity comparing seasons on Pelee Island, ON, using spatially filtered localities and a land cover raster grid using resistance values based on Compton et al. 2007 (see Table S1). Left panel: Connectivity based on *Ambystoma* salamander locations from March and April 2015-2019 (*n* = 190). Center panel: Connectivity based on *Ambystoma* salamander locations from May – July 2015-2019 ($n = 233$). Right panel: Connectivity based on *Ambystoma* salamander locations from August and September 2015-2019 (*n* = 136). Green indicates well-connected areas, with orange and red indicating areas with low connectivity.

Figure S5: Effect of distance to breeding site (m) on predicted habitat suitability for adult and juvenile life stages of *Ambystoma* salamanders on Pelee Island, ON, Canada collected from 2015- 2019. Dashed line depicts the relationship for adult *Ambystoma* salamanders. Solid line depicts the relationship for juvenile *Ambystoma* salamanders. Higher values on y-axis represent higher habitat suitability.

Figure S6: Effect of distance to breeding site (m) on predicted habitat suitability *Ambystoma* salamanders on Pelee Island, ON, Canada across seasons using data collected from 2015-2019. Dotted line indicates model response based on *Ambystoma* salamander locations from March-April (*n* = 190). Dashed line indicates model response based on *Ambystoma* salamander locations from May-July (*n* = 233). Solid line indicates model response based on *Ambystoma* salamander locations from August-September ($n = 136$). Higher values on y-axis represent higher habitat suitability.

Figure S7: Effect of tested environmental variables on predicted habitat suitability for *Ambystoma* salamanders (*n* = 476) on Pelee Island, ON, Canada collected from 2015-2019. A) Effect of land cover classifications on Maxent model. Land cover classes were 1: Wetland, 2: Forest, 3: Tallgrass Community, 4: Agricultural, 5: Transportation, 6: Human Disturbance, 7: Open Water. B) Effect of elevation on Maxent model. C) Effect of soil type on Maxent model. Soil classes were 1: Eastport Sand, 2: Farmington Loam, 3: Brookston Clay, 4: Toledo Clay, 5: Marsh. Higher values on y-axis represent higher habitat suitability.

Figure S8: Effect of land cover type on predicted habitat suitability of *Ambystoma* salamanders on Pelee Island, ON, Canada collected from 2015-2018, based on tested genomotypes. A) locations of *Ambystoma texanum* (TT) individuals (*n* = 37). B) locations of unisexual diploid (LT) individuals (*n* = 286). C) locations of unisexual triploid (LTT) individuals (*n* = 156). Land cover classes were 1 – Wetland, 2 – Forest, 3 – Tallgrass Community, 4 – Agricultural, 5 –

Transportation, 6 – Human Disturbance, 7 – Open Water. Higher values on y-axis represent higher habitat suitability. Note the different scales on the y-axis.

Figure S9: Effect of soil type on predicted habitat suitability of *Ambystoma texanum* (TT) salamanders on Pelee Island, ON, Canada collected from 2015-2018. Soil type classes were 1- Eastport Sand, 2- Farmington Loam, 3- Brookston Clay, 4- Toledo Clay, 5- Marsh. Higher values on y-axis represent higher habitat suitability.

Figure S10: Effect of land cover type on predicted habitat suitability of *Ambystoma* salamanders on Pelee Island, ON, Canada collected from 2015-2019, based on tested life stages. A) locations of adult salamanders ($n = 369$). B) locations of juvenile salamanders ($n = 179$). Land cover classes were 1: Wetland, 2: Forest, 3: Tallgrass Community, 4: Agricultural, 5: Transportation, 6: Human Disturbance, 7: Open Water. Higher values on y-axis represent higher habitat suitability.

Figure S11: Effect of land cover type on predicted habitat suitability of *Ambystoma* salamanders on Pelee Island, ON, Canada collected from 2015-2019, based on tested seasons. A) locations collected in March and April ($n = 190$). B) locations collected from May-July ($n = 233$). C) locations collected in August and September $(n = 136)$. Land cover classes were $1 -$ Wetland, $2 -$ Forest, 3 – Tallgrass Community, 4 – Agricultural, 5 – Transportation, 6 – Human Disturbance, 7 – Open Water. Higher values on y-axis represent higher habitat suitability. Note the different scales on the y-axis.

Figure S12: Distribution of habitat suitability of *Ambystoma* salamanders across Pelee Island, ON, Canada collected from 2015-2019, based on predicted suitability of individual 10x10m grid cells ($n = 173,764$) from model output using all salamander location data ($n = 476$). Suitability values ranged from 0 to 1: values <0.2 represented unsuitable habitat, values between 0.2 and 0.4 represented low suitability, values between 0.4 and 0.6 represented intermediate suitability and values >0.6 represented high suitability.

Figure S13: Distribution of habitat suitability of *Ambystoma* salamanders across Pelee Island, ON, Canada collected from 2015-2018, based on predicted suitability of individual 10x10m grid cells (*n* =173,764) from model outputs of tested genomotypes. Black represents all unisexual diploid (LT) locations ($n = 286$); Dark gray represents all unisexual triploid (LTT) locations ($n =$ 156); Light gray represents all *Ambystoma texanum* (TT) locations (*n* = 37). Suitability values ranged from 0 to 1: values <0.2 represented unsuitable habitat, values between 0.2 and 0.4 represented low suitability, values between 0.4 and 0.6 represented intermediate suitability and values >0.6 represented high suitability.

Figure S14: Distribution of habitat suitability of *Ambystoma* salamanders across Pelee Island, ON, Canada collected from 2015-2019, based on predicted suitability of individual 10x10m grid cells (*n* =173,764) from model outputs of tested life stages. Black represents all adult salamander location data ($n = 369$); Gray represents all juvenile salamander location data ($n = 179$). Suitability values ranged from 0 to 1: values <0.2 represented unsuitable habitat, values between 0.2 and 0.4 represented low suitability, values between 0.4 and 0.6 represented intermediate suitability and values > 0.6 represented high suitability.

Figure S15: Distribution of habitat suitability of *Ambystoma* salamanders across Pelee Island, ON, Canada collected from 2015-2019, based on predicted suitability of individual 10x10m grid cells (*n* =173,764) from model outputs of tested seasons. Black represents all locations collected in March and April ($n = 190$); Dark gray represents all locations collected from May-July ($n =$ 233); Light gray represents all locations collected in August and September ($n = 136$). Suitability

values ranged from 0 to 1: values <0.2 represented unsuitable habitat, values between 0.2 and 0.4 represented low suitability, values between 0.4 and 0.6 represented intermediate suitability and values >0.6 represented high suitability.

List of Tables

Table 1: Patch-level, class-level and landscape-level metrics measured with Fragstats for highly suitable habitat for *Ambystoma texanum* and unisexual Ambystoma on Pelee Island, ON. All metrics were measured at the class-level, aside from AREA and PERIM measured at patch-level and GYRATE measured at landscape-level. See main text for metric descriptions. The model with all locations was created with data from 2015-2019, and genotype-specific models were created with data from 2015-2018.

Table 2: Percent contribution and permutation importance of selected environmental variables in Maxent models for *A. texanum* and unisexual *Ambystoma* captured from 2015 – 2019, and from 2015-2018 for genotype-specific models.

Table S1: Resistance values created for *Ambystoma* salamanders on Pelee Island, ON, Canada, based on work by Compton et al. (2007). A resistance value of 1 indicates minimal resistance (i.e., most easy to travel through), and a value of 20 indicates maximum resistance (i.e., movement completely impeded). This table also outlines how we grouped land cover types in our analyses to reduce the dimensionality of this variable.

Table S2: Proportion of land cover types across Pelee Island, ON, according to the classification used in habitat suitability models.

Table S3: AUC values of Maxent models for *Ambystoma texanum* and unisexual *Ambystoma*. Training data AUC is calculated from training data selected from the data set, and test data AUC is the real test of the model's predictive power.

Chapter 1: General Introduction

Habitat loss and fragmentation are among the main causes of ecosystem disruption, including eliciting changes in the distribution and abundance of native plants and animals (Mantyka‐Pringle et al. 2012; Haddad et al. 2015). Loss of available habitat, particularly through the fragmentation of larger habitat patches into small areas with low connectivity to other suitable areas, is detrimental for the persistence of many populations and metapopulations (Debinski and Holt 2000). Contemporary habitat loss and fragmentation are primarily caused by anthropogenic disturbances, including the conversion of natural ecosystems to agricultural and urban development and the construction of roadways and other geographic barriers that limit dispersal and genetic exchange (Hoekstra et al. 2005; Coristine and Kerr 2011). Impacts of habitat loss and fragmentation are aggravated in species with restrictive habitat requirements or that have low mobility or dispersal ability (Vos et al. 2002), and it follows that modelling the potential spatial distribution of such species across a range of habitat loss and fragmentation scenarios is crucial for prioritizing conservation planning efforts targeting habitat protection and restoration.

Effects of Habitat Fragmentation and Loss on Species Populations

Fragmentation refers not only to habitat loss but habitat reconfiguration, where patches of suitable habitat become smaller, more numerous, and increasingly isolated (Fahrig 2003). It is debatable whether the complete loss of habitat is substantially more detrimental to the persistence of species than is habitat fragmentation per se, as fragmentation does not always lead to an immediate decline in distribution and abundance depending on species' habitat requirements and the intensity of habitat change (Jackson and Fahrig 2013; Fletcher et al. 2018). However, habitat fragmentation is still a threat for species whose populations are small and isolated, and these responses may be compounded in light of edge effects that further erode the integrity of marginal habitat (Laurance et al. 2007; Jackson and Fahrig 2013). Accordingly, there can be both direct and indirect effects of habitat change on species, and it remains challenging to disentangle which factors are driving variation, especially when responses are gradual and difficult to track.

Reduced inter-patch movement is characteristic of species exposed to high fragmentation (Debinski and Holt 2000; Haddad et al. 2015), but such impacts can be mitigated when traversable habitat remains available between patches (Vos et al. 2002; Hilty et al. 2019). Continuous corridors are one type of pathway that provides an unbroken stretch of habitat between patches; vegetated overpasses and riparian corridors are examples of continuous corridors that may be used by a variety of wild animal species ranging from mammals to amphibians (Hilty et al. 2019). As a rule of thumb, corridors should constitute the shortest length possible between suitable habitat patches and be comprised of high-quality habitat. However, it is not always possible to establish short, continuous high-quality corridors between habitat patches, meaning that stepping-stone corridors comprised of lesser-quality habitat may be necessary (Lindenmayer and Fischer 2006). These corridors would primarily benefit species that can move rapidly through intermediate habitat (Doeer et al. 2010), and are known to have successfully linked habitat patches for species such as kiwis (e.g., *Apteryx mantelli*; Potter 1990) and some butterflies (e.g., *Proclassiana eunomia*; Neve et al. 1996). Yet, for many species with restricted movement or high vulnerability to extirpation when in poor habitat conditions,

lower-quality corridors may be of limited use and populations may become functionally isolated (Henein and Merriam 1990; Gustafsson and Hansson 1997). Given the crucial role that corridors play in maintaining dispersal and breeding opportunities and thus promoting genetic diversity and favourable community dynamics (Schlaepfer et al. 2018), their suitability and functionality should be assessed (Hilty et al. 2019).

Habitat and Connectivity Modelling

Evaluating habitat availability and suitability for species can help support robust conservation planning decisions, including identifying areas that should be prioritized for protection to ensure species persistence (Lindenmayer and Fischer 2006; Hilty et al. 2019). Habitat suitability models support the development of maps outlining the geographic extent of favourable and unfavourable abiotic and biotic variables, using prior observations across similar environmental conditions as the basis for model training (Phillips et al. 2006). In general, habitat suitability models can be designed specifically to assess and identify priority environmental features playing a crucial role in current and potentially future species occurrence and persistence (Elith et al. 2006); the role of such modelling exercises may be especially important for rare or endangered species, those with specific habitat requirements, or identifying possible areas to protect and conserve for the benefit of declining populations (Sattler et al. 2007; Crawford et al. 2020). Numerous studies have modelled future habitat suitability and species distribution, such as evaluating changes in distributions for the Chinese giant salamander (*Andrias davidianus*) – a loss of at least two-thirds of their suitable habitat by 2070 – and the wood turtle (*Glyptemys insculpta*) – a loss of over half of their suitable habitat in the northeastern US by 2070 (Zhang et al. 2020; Mothes et al. 2020). During habitat

3

suitability model development, variables should be chosen based on factors relevant to the environmental niche of the species, which refers to the environmental factors promoting population persistence (Hirzel and Le Lay 2008). There are many recent technological developments that have improved habitat suitability modelling, including remote sensing datasets that provide spatial and temporal information on environmental features relevant to organisms, such as temperature/precipitation, land cover type, and urban activity (Hirzel and Le Lay 2008). It follows that integrating such sources of information into habitat suitability models for at-risk species may play an indispensable role in supporting an informed assessment of the abundance, spatial configuration, and composition of occupiable habitat in an area. Likewise, habitat connectivity or landscape resistance models are valuable tools for assessing existing and potential future linkages between suitable habitat patches; these models evaluate how continuous or resistant an environment is to dispersal based on land cover types (Rudnick et al. 2012). Measuring connectivity allows conservation planners to make informed decisions about whether sub-populations are likely to be facing isolation and what measures may remedy weak linkages to core populations (Bennett 2003).

Study System: Ambystoma *salamanders on Pelee Island*

I evaluated habitat suitability and connectivity of *Ambystoma* salamanders on Pelee Island, Ontario in the extreme southwestern portion of the province of Ontario, in Lake Erie. Salamanders on Pelee Island are currently reliant on a small number of habitat patches, with apparently limited connectivity between suitable patches (Hossie 2018). This group of salamanders consists of small-mouthed salamanders (*Ambystoma texanum*), blue-spotted salamanders (*A. laterale*) and Unisexual *Ambystoma* (COSEWIC

4

2017). Unisexual *Ambystoma* are an all-female lineage, who use sperm packets laid by parental *Ambystoma* species to stimulate egg development; this genetic material can sometimes be incorporated in egg fertilization but more often eggs are clones of their mother and sperm is not incorporated into the egg (Bogart et al. 2007). Five *Ambystoma* salamander species throughout the Great Lakes region in eastern North America can promote egg development in unisexuals, but only *A. texanum* and *A. laterale* are present on Pelee Island (Bogart et al. 2007). *Ambystoma* salamanders rely on fishless wetlands to lay eggs in early spring, but while larvae require wetlands for survival, they metamorphose and occupy more terrestrial habitat during mid-late summer (Harding and Mifsud 2017).

Historically, Pelee Island was mostly forested wetlands and included several distinct land bodies separated by sandbars that likely served as barriers to salamander dispersal (Forbes et al. 2000). In the 1880s, these wetlands were deforested and drained to support agriculture, thereby decreasing the size of habitat areas and potentially altering habitat availability for salamanders, as well as the connectivity between terrestrial habitat patches (Forbes et al. 2000; Hossie 2018). In the past 2 decades, conservation organizations and local landowners working on Pelee Island have initiated protection and restoration of natural areas, but nevertheless, salamander habitat remains largely disjunct across the island. This is a concern for a number of reasons, including that: 1) smallmouthed salamanders are listed as endangered in Canada and Ontario and their distribution is entirely restricted to Pelee Island; 2) blue-spotted salamanders, although widely distributed across eastern Canada, are uncommon on Pelee Island and may have distinct genetic status on Pelee Island and thus require special protection (Hossie 2018;

Hossie 2021, unsolicited report); 3) unisexual *Ambystoma* on Pelee Island, although seemingly common on the island, are also listed as endangered in Canada (under the Committee on the Status of Endangered Wildlife in Canada [COSEWIC]) and Ontario and their sustainability is entirely reliant on the viability of parental species that serve as sperm donors (small-mouthed and blue-spotted salamanders). It follows that there is an urgent need to assess the distribution and quality of both terrestrial and aquatic salamander habitat on Pelee Island, to determine requirements for viability and persistence of this assemblage.

Research Questions

The aim of this thesis is to analyze salamander habitat suitability and connectivity in a fragmented landscape on Pelee Island. I addressed this research objective by modelling habitat suitability and connectivity using location data for Pelee Island salamanders collected over 5 years (2015-2019). The extensive sampling effort yielded larval, juvenile, and adult *Ambystoma* salamanders from across the assemblage of groups, and salamander capture methods included coverboards, minnow traps, and dipnet surveys. In 2019, I added additional coverboards specifically to sample under-surveyed areas, including locations expected to have a low probability of salamander occurrence. Models of salamander habitat suitability and connectivity were supported by geospatial environmental data representing the primary landcover and abiotic factors that were likely to influence salamander distribution and abundance. The research questions addressed in this thesis are:

• How much habitat is available on Pelee Island for the *Ambystoma* complex? Historical forested regions and populations of salamanders have been fragmented

6

by increased agricultural and urban development, meaning that suitable habitat is expected to be patchily distributed across the landscape.

- How connected are the remaining patches of suitable salamander habitat? Salamander assemblages should have some level of connectivity between occupied habitat patches to maintain dispersal and genetic exchange, but patches may be too isolated if they are beyond the normal dispersal distance of *Ambystoma*.
- What is the state of remaining habitat for small-mouthed salamanders specifically, and how connected are their populations? Small-mouthed salamanders are likely the sole sperm donor for unisexuals in many breeding locations, and thus are disproportionally important to the complex. Due to their higher habitat specificity and tendency to remain close to breeding sites compared to unisexuals, smallmouthed salamanders likely have a smaller amount of suitable habitat on Pelee Island as compared to unisexual *Ambystoma*. In contrast, unisexuals are more abundant than small-mouthed salamanders and likely tolerate a wider range of habitats due to the varied composition of their nuclear genome (Mills et al. 2020).

References

- Bennett AF (2003) Linkages in the Landscape: The Role of Corridors and Connectivity in Wildlife Conservation, 2nd edn. IUCN, Gland, Switzerland and Cambridge, UK
- Bogart J, Bi K, Fu J, et al (2007) Unisexual salamanders (genus Ambystoma) present a new reproductive mode for eukaryotes. Genome 50:119–136. https://doi.org/10.1139/G06-152
- Coristine LE, Kerr JT (2011) Habitat loss, climate change, and emerging conservation challenges in Canada. Can J Zool 89:435–451. https://doi.org/10.1139/z11-023

Committee on the Status of Endangered Wildlife in Canada (COSEWIC) (2017) Unisexual ambystoma (Ambystoma laterale) and 3 subspecies: COSEWIC assessment and status report 2016. In: Gov. Can. https://www.canada.ca/en/environment-climate-change/services/species-riskpublic-registry/cosewic-assessments-status-reports/unisexual-ambystomasubspecies-2016.html. Accessed 6 Jan 2021

- Crawford BA, Maerz JC, Moore CT (2020) Expert-Informed Habitat Suitability Analysis for At-Risk Species Assessment and Conservation Planning. J Fish Wildl Manag 11:130–150. https://doi.org/10.3996/092019-JFWM-075
- Debinski DM, Holt RD (2000) A Survey and Overview of Habitat Fragmentation Experiments. Conserv Biol 14:342–355. https://doi.org/10.1046/j.1523- 1739.2000.98081.x
- Doeer VAJ, Doeer ED, Davies MJ (2010) Does structural connectivity facilitate dispersal of native species in Australia's fragmented terrestrial landscapes. Collaboration for Environmental Evidence, Canberra, AUS
- Elith J, Graham CH, Anderson RP, et al (2006) Novel methods improve prediction of species' distributions from occurrence data. Ecography 29:129–151. https://doi.org/10.1111/j.2006.0906-7590.04596.x
- Fahrig L (2003) Effects of Habitat Fragmentation on Biodiversity. Annu Rev Ecol Evol Syst 34:487–515. https://doi.org/10.1146/annurev.ecolsys.34.011802.132419
- Fletcher RJ, Didham RK, Banks-Leite C, et al (2018) Is habitat fragmentation good for biodiversity? Biol Conserv 226:9–15. https://doi.org/10.1016/j.biocon.2018.07.022
- Forbes MR, Schalk G, Hawco SM, Prior KA (2000) Ecosystem Plan for Pelee Island. In: Proceedings of a Conference on the Biology and Management of Species and Habitats at Risk. B.C. Ministry of Environment, Lands and Parks, Victoria, B.C. and University College of the Cariboo, Kamloops, B.C, Kamloops, BC, pp 903– 908

Gustafsson L, Hansson L (1997) Corridors as a Conservation Tool. Ecol Bull 182–190

- Haddad NM, Brudvig LA, Clobert J, et al (2015) Habitat fragmentation and its lasting impact on Earth's ecosystems. Sci Adv 1:e1500052. https://doi.org/10.1126/sciadv.1500052
- Harding JH, Mifsud DA (2017) Amphibians and Reptiles of the Great Lakes Region, Revised Ed. University of Michigan Press, Ann Arbor, Michigan
- Henein K, Merriam G (1990) The elements of connectivity where corridor quality is variable. Landsc Ecol 4:157–170
- Hilty J A, Keeley ATH, Lidicker WZ, Merelender AM (2019) Corridor Ecology, Second Edition: Linking Landscapes for Biodiversity Conservation and Climate Adaptation, 2nd edn. Island Press, Washington, DC.
- Hirzel AH, Le Lay G (2008) Habitat suitability modelling and niche theory. J Appl Ecol 45:1372–1381. https://doi.org/10.1111/j.1365-2664.2008.01524.x
- Hoekstra JM, Boucher TM, Ricketts TH, Roberts C (2005) Confronting a biome crisis: global disparities of habitat loss and protection. Ecol Lett 8:23–29.
- Hossie TJ (2018) Recovery Strategy for Small-mouthed Salamander (Ambystoma texanum) and Unisexual Ambystoma Small-mouthed Salamander dependent population (Ambystoma laterale - texanum) in Ontario. Ontario Ministry of Natural Resources and Forestry, Peterborough, Ontario.
- Jackson HB, Fahrig L (2013) Habitat Loss and Fragmentation. In: Levin SA (ed) Encyclopedia of Biodiversity (Second Edition). Academic Press, Waltham, pp 50–58
- Laurance WF, Nascimento HEM, Laurance SG, et al (2007) Habitat Fragmentation, Variable Edge Effects, and the Landscape-Divergence Hypothesis. PLOS ONE 2:e1017. https://doi.org/10.1371/journal.pone.0001017
- Lindenmayer DB, Fischer J (2006) Habitat Fragmentation and Landscape Change: An Ecological and Conservation Synthesis. Island Press, Washington, DC.
- Mantyka‐Pringle CS, Martin TG, Rhodes JR (2012) Interactions between climate and habitat loss effects on biodiversity: a systematic review and meta-analysis. Glob Change Biol 18:1239–1252. https://doi.org/10.1111/j.1365-2486.2011.02593.x
- Mills PB, Hossie TJ, Murray DL (2020) Niche determinants in a salamander complex: Does hybridism or reproductive parasitism explain patterns of distribution? Ecosphere 11:e03265. https://doi.org/10.1002/ecs2.3265
- Mothes CC, Howell HJ, Searcy CA (2020) Habitat suitability models for the imperiled wood turtle (Glyptemys insculpta) raise concerns for the species' persistence

under future climate change. Glob Ecol Conserv 24:e01247. https://doi.org/10.1016/j.gecco.2020.e01247

- Neve G, Barascud B, Hughes R, et al (1996) Dispersal, Colonization Power and Metapopulation Structure in the Vulnerable Butterfly Proclossiana eunomia (Lepidoptera: Nymphalidae). Journal of Applied Ecology 33:14–22. https://doi.org/10.2307/2405011
- Phillips SJ, Anderson RP, Schapire RE (2006) Maximum entropy modeling of species geographic distributions. Ecological Modelling 190:231–259. https://doi.org/10.1016/j.ecolmodel.2005.03.026
- Potter MA (1990) Movement of North Island brown kiwi (Apteryx australis mantelli) between forest fragments. New Zealand Journal of Ecology 14:8
- Rudnick DA, Ryan SJ, Beier P, et al (2012) The role of landscape connectivity in planning and implementing conservation and restoration priorities. Ecological Society of America, Rocky Mountain Research Station, Fort Collins, CO
- Sattler T, Bontadina F, Hirzel AH, Arlettaz R (2007) Ecological niche modelling of two cryptic bat species calls for a reassessment of their conservation status. J Appl Ecol 44:1188–1199. https://doi.org/10.1111/j.1365-2664.2007.01328.x
- Schlaepfer DR, Braschler B, Rusterholz H-P, Baur B (2018) Genetic effects of anthropogenic habitat fragmentation on remnant animal and plant populations: a meta-analysis. Ecosphere 9:e02488. https://doi.org/10.1002/ecs2.2488
- Vos CC, Baveco H, Grashof-Bokdam CJ (2002) Corridors and Species Dispersal. In: Gutzwiller KJ (ed) Applying Landscape Ecology in Biological Conservation. Springer New York, New York, NY, pp 84–104
- Zhang Z, Mammola S, Liang Z, et al (2020) Future climate change will severely reduce habitat suitability of the Critically Endangered Chinese giant salamander. Freshw Biol 65:971–980. https://doi.org/10.1111/fwb.13483

Chapter 2: Assessing habitat suitability and connectivity for an

endangered salamander complex

Graeme N. SMITH¹, Evan A. BARE¹, Thomas J. HOSSIE², Dennis L. MURRAY²

- ¹ Environmental and Life Sciences Graduate Program, Trent University, 2140 East Bank Drive, Peterborough, ON, Canada K9J 7B8
- ² Department of Biology, Trent University, 2140 East Bank Drive, Peterborough, ON, Canada K9J 7B8

Abstract:

Context

Evaluating habitat quality for isolated animal communities is a high priority in landscape ecology and conservation biology. Where possible, such efforts should focus on understanding the needs of species that are disproportionately important for maintaining diversity.

Objectives

We evaluated the habitat of *Ambystoma* salamanders on Pelee Island, ON, Canada,

including unisexuals and the bisexual species on which they are reproductively dependent

(*A. texanum*). Existing assemblages on the island appear isolated from one another,

perhaps reflecting limited habitat suitability and low connectivity.

Methods

Field surveys resulted in 2076 salamander detections in terrestrial and aquatic areas. We modelled habitat suitability using MaxEnt and connectivity with Circuitscape.

Results

Island-wide habitat availability was low (6.9%). Proximity to wetlands was crucial for habitat suitability, with forested areas and non-breeding wetlands being suitable only when found near established breeding locations. *A. texanum*, the critical sperm donor species, had 44.8% less suitable habitat area compared to its unisexual dependents. On average, existing salamander assemblages were separated by 2.1 ± 1.3 (SE) km, which is beyond the typical dispersal distance for *Ambystoma*. Potential dispersal corridors were primarily comprised of unsuitable habitat for salamanders.

Conclusions

The *Ambystoma* salamander complex on Pelee Island hinges on the viability of a sperm donor species with marked habitat and dispersal restrictions. Combined with contemporary anthropogenic land use, this has created isolated assemblages with uncertain long-term viability. Preserving this complex will require habitat protection in proximity to established breeding locations, as well as the enhancement of dispersal corridors to increase connectivity among patches of suitable habitat.

Keywords: Amphibians, landscape fragmentation, habitat suitability, connectivity, landscape resistance, unisexual

Introduction:

Habitat loss and fragmentation are among the primary sources of biodiversity loss globally (Thomas et al. 2004; Hoffmann et al. 2010). Land conversion has reduced the total amount of viable habitat and reduced connectivity between increasingly isolated habitat patches, resulting in population declines, species loss, and disruption of community structure and function (Mantyka‐Pringle et al. 2012; Haddad et al. 2015). These impacts are particularly detrimental for species with strong habitat specialization, small areas of occupancy, and low mobility (Fischer and Lindenmayer 2007). It follows that when species have multiple life stages with distinct environmental needs, habitatrelated challenges to population persistence may be especially harmful (Becker et al. 2007). In fragmented landscapes, individuals in legacy habitat patches may persist as components of a metapopulation if sub-populations are sufficiently connected to adjacent populations; without such connections, sub-populations experience a higher risk of local extinction (Hanski 1998). Ultimately, metapopulation persistence depends on having functional connectivity among habitat patches and meeting minimum requirements of habitat patch size and suitability (Keymer et al. 2000).

The above principles of metapopulation dynamics extend to ecological communities and assemblages, where persistence at a site requires meeting the habitat needs of a variety of members. Further, the habitat requirements of some species can disproportionately influence the composition and viability of communities or assemblages when their membership is highly inter-dependent and contingent on the presence of other species (Callaway 2007; Hatcher et al. 2012). In such cases, entire communities may be especially vulnerable when key members have restricted habitat

requirements or dispersal ability, and community persistence may critically depend on habitat that favours these species. Further, maintaining metapopulation dynamics requires that habitat patches are well-connected through immigration/emigration and recolonization, especially to preserve genetic diversity (Crooks and Sanjayan 2006). Identifying potential corridors is central to determining whether functional metapopulations exist or whether metapopulation dynamics can be restored in fragmented landscapes (Rudnick et al. 2012; Dondina et al. 2018). Functionally, effective corridors linking suitable habitat patches can range from poor-quality areas with a low resistance to movement, to high-quality habitat patches that constitute suitable habitat (Beier and Noss 1998; Bennett 2003). Importantly, connectivity metrics do not guarantee functional connectivity, and landscape resistance modelling can provide a more holistic view of the potential connectivity within a landscape (Bennett 2003; Pierik et al. 2016).

In contemporary landscapes, habitat loss and fragmentation are particularly impactful to forest and wetland systems (Mantyka‐Pringle et al. 2012). Wetlands have experienced especially dramatic declines during the last century (Davidson 2014; Dixon et al. 2016), including in southern Ontario, Canada, where >85% of original wetlands have been lost (Ducks Unlimited Canada 2010; Environmental Commissioner of Ontario 2018). Many amphibians depend on wetlands as critical habitat for egg-laying and larval development. In addition, amphibians often require suitable terrestrial habitat because of their biphasic life history, meaning that comprehensive suitability assessments should consider both wetlands and nearby terrestrial habitat (Hecnar 2004; Scott et al. 2013). Declining landscape connectivity as a result of habitat fragmentation remains an

important cause of amphibian declines and can be especially severe when terrestrial and aquatic habitats become decoupled (Lehtinen et al. 1999).

Canada's only population of small-mouthed salamanders (*Ambystoma texanum*) occurs on Pelee Island, Ontario, where they live alongside blue-spotted salamanders (*A. laterale*) and unisexual *Ambystoma* salamanders (Small-mouthed salamander dependent population) (Bogart et al. 1985; Hossie 2018). Unisexual *Ambystoma* are an all-female lineage that are reproductively-dependent on sperm produced by males of other *Ambystoma* species, although they all possess at least one *A. laterale* genome (Bogart et al. 2007). Unisexuals typically produce clonal offspring, but can also produce ploidyelevated offspring when the nuclear DNA from the sperm is incorporated into the egg (Bogart and Licht 1986; Bogart 2019b). *A. texanum* are disproportionately important to this *Ambystoma* complex, as the few remaining *A. laterale* have a highly constrained distribution on Pelee Island, and unisexual *Ambystoma* are almost exclusively reliant on *A. texanum* to reproduce (Hossie 2018). To complete their larval phase, all three of these *Ambystoma* salamanders require fishless breeding locations in proximity to forested terrestrial habitat (Porej et al. 2004; Porej and Hetherington 2005). Adult *A. texanum* salamanders do not travel far from their breeding locations (Gopurenko et al. 2007), although they are capable of moving >150 m in a single movement session (Williams 1973; Denton et al. 2017) across suitable habitat. By contrast, unisexuals travel <50 m in a single session (Denton et al. 2017), but may tolerate a wider range of habitats (Greenwald et al. 2016). This can impact the metapopulation structure of the salamander complex if sites are isolated and dispersal is impeded by barriers like agriculture or human development that impact the three salamander taxa differently (Marsh and

15

Trenham 2001; Greenwald et al. 2016). Further, characterizing differences in habitat suitability between host species and reproductively-dependent unisexuals is essential for conserving this salamander complex.

We assessed suitability and connectivity of *Ambystoma* salamander habitat on Pelee Island in southern Ontario and predicted that limited natural habitat and extensive fragmentation have caused disjunct salamander populations rather than a functional metapopulation. Specifically, we predicted that remaining habitat patches would be structured in relatively small patches that are separated by a distance exceeding the typical dispersal distance for *Ambystoma* salamanders. We expected that *A. texanum* – which are a key member of the *Ambystoma* salamander complex on Pelee Island but have low overall availability relative to unisexuals – would have higher habitat specificity, including restricted proximity to breeding locations and lesser suitability of potential corridors, resulting in lower overall habitat availability compared to unisexuals. On Pelee Island, *A. texanum* are the more abundant and widely distributed host compared to *A. laterale* (Bare et al. *in review*), and therefore they are central to supporting the *Ambystoma* complex in our study area and thus are the focal species for our investigation. Finally, we also predicted that unisexuals should generally tolerate a wider range of habitats and longer dispersal distances than *A. texanum*. However, because unisexuals can take on a variety of genetic forms (i.e., genomotypes), their habitat suitability and connectivity requirements should more closely reflect those of *A. texanum* depending on the number of *A. texanum* genomes present in individual animals.

16

Methods:

Study Area and System

Our study was conducted on Pelee Island, Ontario, Canada (41.77 N, 82.66 W), a 42 km² island in Lake Erie that holds Canada's only population of *A. texanum* (Hossie, 2018). Historically, the island was heavily forested and covered with wetlands, but currently forests and wetlands only cover 16.5% and 5.7% of the island, respectively (G. Smith, unpublished data). Agricultural land is now the primary land cover type (62.1%), with remnant forest and wetlands scattered across the island. Unisexual *Ambystoma* (*Ambystoma laterale - texanum*) are reproductive parasites and possess nuclear DNA from both *A. texanum* and *A. laterale* hosts (Bogart et al. 2009; COSEWIC 2017). A unique nomenclature is used to distinguish among the various unisexual forms ("genomotypes") that can arise based on the number of *A. laterale* ('L') and *A. texanum* ('T') chromosome sets they possess (Lowcock et al. 1987; Bogart et al. 2007). For example, a diploid unisexual with a single chromosome set from *A. laterale* and *A. texanum* would be referred to as 'LT' and a *texanum*-dominant triploid would be 'LTT'. Salamander identity was determined by genotyping tissue samples using microsatellite loci (*Atex74, Atex102, Atex141, AjeD75, AjeD94, AjeD283, AjeD346, AjeD378 AjeD422*) (see also Williams and DeWoody 2004; Peterman et al. 2012), and these methods have been fully detailed elsewhere (Bogart 2019a; Van Drunen et al. 2020).

A. texanum use several types of moist habitats, including tall-grass prairies, dense hardwood forests, and occasionally agricultural land when it is adjacent to suitable breeding locations (Environment and Climate Change Canada 2020). Adults frequently use burrows created by semi-terrestrial crayfish (Williams 1973; Owen and Jutterbock

2013). Microhabitat requirements also differ between groups, with *A. texanum* preferring moist, forested habitat and *A. laterale* being more tolerant of drier soils and open habitat (Jutterbock and Owen 2013; Mills et al. 2020). However, it is not clear whether these different preferences lead to landscape-level differences in habitat requirements across the salamander complex. Unisexual *Ambystoma* with genomes from *A. texanum* may be restricted to areas with clay-based soils (Petranka 1998), but recent work shows that the 'hybrid' nuclear genome of unisexual *Ambystoma* gives rise to intermediate habitat requirements compared to their hosts (Mills et al. 2020). It follows that unisexual *Ambystoma* should occupy intermediate habitat to their sperm-donor host species (see Greenwald et al. 2016; Mills et al. 2020).

Fieldwork

During March-August (2015-2019), monthly salamander surveys were conducted at various sites across the island $(n = 10)$ (with additional sampling in September 2019). Here, we define 'site' as a large area of suitable habitat (including both terrestrial and aquatic landcover) separated from other areas of suitable habitat by more than the typical dispersal distance for *Ambystoma* salamanders (i.e., > 600 m, acknowledging that infrequent dispersal up to 750-1350 m has been reported: Madison 1997; Gamble et al. 2007). These sites were all separated by agricultural land and urban development, and range in size from 16 to 240 ha (and some sites span multiple protected areas) (G. Smith, unpublished data). Sites contain breeding locations, which we define as bodies of water with evidence of salamander breeding; multiple breeding locations can be present at a given site. Over this time from 2015-2019, salamanders were captured through opportunistic searches, systematic searches using coverboards, and minnow-trapping for

adults during the breeding season (March). Coverboard arrays were established between 2015-2017 at all sites across the island with known salamander populations ($n = 304$) boards). We identified breeding locations by minnow-trapping breeding adults, conducting visual egg searches, and dip-netting for larvae. The only confirmed breeding location with *A. laterale* was not sampled as it was on private land. In May and June 2019, an additional 157 coverboards were deployed strategically across the island to improve spatial coverage of survey efforts, including areas predicted to have low salamander occupancy rates. Based on preliminary models using data from 2015-2018, 106 boards were placed in areas predicted to be movement areas and corridors (i.e., with a high current flow as per Circuitscape models, see below) and were separated by at least 50 m; the remaining 51 boards were spaced 300 m apart and placed in random locations with lower predicted flow. For all 461 boards, most (57.1%) were in forested areas, with the rest in wetlands, tallgrass communities and agricultural land within protected areas (Proportion of boards in forested areas: 57.1%; wetlands: 22.3%, tallgrass comm.: 15.2%, agricultural: 5.4%). From June - September 2019, we conducted three surveys of the new boards. We also searched natural cover objects within 5 m of boards, including at the time of deployment, to further increase salamander detection. To supplement these efforts, linear transects that involved overturning natural cover objects were conducted in areas predicted to have low and high habitat connectivity (based on preliminary models), from June - September 2019 (June: *n* = 19, August: *n* = 29, September: *n* = 34). During these transect searches, three observers spaced 2.5 m apart walked 50 m transects. Finally, we also conducted opportunistic searches in areas known to have salamanders. We combined capture data from all sampling periods (i.e., 2015-2019) for our analyses,

although genetic data was not completed for the samples collected in 2019 so genotypespecific analyses were restricted to data from only 2015-2018.

Habitat Suitability

We used Maxent (version 3.4.0) to assess salamander habitat suitability on the island. Maxent combines organism presence data with environmental grids to generate a predicted distribution for the species of interest (Elith et al. 2011). Salamander locations (*n* = 1237), where 'location' refers to the specific geographic location where a salamander was found, were used to develop the preliminary Maxent model, with 40% of locations used in model training. Location points were spatially clustered, so we spatially filtered our data to a single observation per 10 x 10 m grid cell, resulting in 476 unique locations (22.9% of the complete dataset). Distance to breeding location, land cover type, soil type, and elevation served as relevant environment variables given that: i) Ambystomatids typically remain close to breeding areas, inhabiting shaded woodland areas with soft, moist soil (Harding and Mifsud 2017); ii) agricultural land is typically avoided by *Ambystoma* spp. (Kolozsvary and Swihart 1999; Greenwald et al. 2009); and iii) elevation may help distinguish areas likely to remain wet vs. becoming dry by late summer. Land cover and soil type were treated as categorical variables, while distance to breeding location and elevation were continuous. Land cover was based on the Southern Ontario Land Resource Information System (SOLRIS) (SOLRIS Technical Team 2015). For 'undifferentiated' land cover types, Google Earth (version 9.2.78.1, accessed February 2019) served to visually classify land cover types. We also collapsed some land cover categories to simplify this variable (see Table S1). Soil data were derived from the Ministry of Natural Resources Land Information Ontario Data Description Soil Survey

Complex (OMAFRA 2019) and elevation data were from the Ministry of Natural Resources Provincial Digital Elevation Model - South (Provincial Mapping Unit 2018). Distance to breeding location was calculated in ArcGIS 10.7.1 based on the distance of each grid cell from nearest margin of a breeding location. Environmental variables were converted from their source file type to rasters, with a cell size of 10 x 10 m.

Probability of suitable habitat values ranged from 0 to 1; here we treated values 0- 0.2 as unsuitable habitat, 0.2 -0.4 as low, 0.4-0.6 as intermediate, and >0.6 as high suitability (see also Atmoko et al., 2020; Singh et al., 2017; Yuh et al., 2020). Habitat suitability was interpreted using median output value of a specified area to compare suitability across various scenarios. Response curves were generated to assess variable fit (Phillips 2017). In these plots the y-axis is given as a logistic output, which represents probability that a species is present and categories within a specific environmental variable are ranked. Variable contribution to model and permutation importance were tested via jackknife. Additional model scenarios were created by subdividing location points to compare salamander genomotype (LT: $n = 286$, LTT: $n = 156$, and TT: $n = 37$), and we also modelled habitat suitability according to salamander life stage (adults: *n* = 369, juveniles: *n* = 179) and seasonality (March and April: *n* = 190, May-July: *n* = 233, and August and September: $n = 136$) to better understand sources of variability in our results. Genomotype and seasonality datasets only consisted of adult capture locations (SVL > 40 mm) and excluded recently-metamorphosed juveniles which emerge as early as late June (all juveniles initially remain close to their natal location). Finally, highly suitable habitat for all *Ambystoma*, LT only, LTT only, and TT only was characterized using Fragstats, a program used to quantify landscape structure, and Patch Analyst, a

Fragstats extension for ArcGIS (Elkie et al. 1999; McGarigal 2015). Variables measured include mean patch area (MPA), perimeter length (PERIM), radius of gyration (GYRATE), perimeter-area fractal dimension (PAFRAC), clumpiness index (CLUMPY), mean nearest neighbour (MNN), number of patches (NP), patch density (PD), and mean proximity index (MPI) (McGarigal 2015).

Habitat Connectivity

Circuitscape (version 4.0) was used to predict ecological connectivity between communities and identify potential movement routes between disconnected areas (McRae et al. 2014). Circuitscape models were created with spatially filtered locations, using the same location points as with Maxent models $(n = 476)$. Landscape features are given a resistance value based on animal travel likelihood through that area (Table S1). Resistance values were based on work by Compton et al. (2007) who conducted a resistant-kernel estimator for *A. opacum* and *A. maculatum* and created resistance values for dispersal from breeding locations and migration (see Table S1). Resistance values were based on land cover types following the same classification used in our previous habitat suitability analyses. Landscape grids were created using ArcGIS 10.7.1 with a cell size of 10 x 10 m to enable the detection of fine-scale movement corridors. Resistance values were interpreted by Circuitscape (in ohms) as electrical resistance, and circuit maps were created between locations points with a 'current' value for each grid cell (in amperes) (McRae et al. 2008). There is no upper range limit for these values, but high output values represent increased likelihood of movement through these areas (McRae et al. 2008). Potential movement corridors linking core sites were identified using Linkage Mapper version 2.0.0 (McRae and Kavanagh 2011), which combines GIS and

Circuitscape to identify potential habitat corridors and pinch-points (where movement is funnelled between core habitat areas) (Gallo and Greene 2018). Linkage Mapper is packaged within ArcGIS and uses a resistance grid and map of core areas. We used the same map of resistance values as with our habitat connectivity models, with breeding locations serving as core areas. Finally, we generated a map of linkages between breeding locations (termed 'core areas') illustrating least-cost paths, which represent the shortest and most likely route an animal would take between patches, which were overlaid on the initial map (Sawyer et al. 2011; Gallo and Greene 2018). All parameters in the model were set to their default setting, creating links between all focal areas and dropping links that intersect focal areas or were >50 km.

Results:

During 2015-19, we caught 2076 post-metamorphic salamanders (2015: 4.4%; 2016: 15.6%; 2017: 19.7%; 2018: 19.9%; 2019: 40.4%) from 10 sites across Pelee Island. Most animals (52.2%) were captured during wetland surveys (of breeding and nonbreeding wetlands), with others captured during opportunistic searches on land under natural cover objects (25.3%), coverboards (18.4%) or on linear transects (2.9%). Most captured animals (83.5%) were adults $(SVL > 40$ mm) with the remainder being newlytransformed individuals (SVL 21-40 mm). Most captures occurred during March-April (67.0%), followed by May-July (19.6%) and August-September (13.5%). Fifty-eight percent of our total sample of adults and juvenile salamanders (*n* = 1197) were genotyped (i.e., all samples collected from 2015-18) and we found that 96% were unisexual *Ambystoma* (LT: 65.3%; LTT: 19.6%; LTTT: 3.8%, LTTTT: 0.08%), with parental species being rare (*A. texanum*: 3.5%; *A. laterale*: 0.2%). Evidence of breeding

(salamander egg masses or larvae) was documented at 33 natural and constructed wetlands (i.e., including vernal pools, forested wetlands, marshes, and swamps) across the island.

Overall Habitat Suitability (All Ambystoma*)*

Our habitat suitability assessment revealed that the majority of Pelee Island is not currently suitable for *Ambystoma* salamanders (93.1% of the island has a Maxent value < 0.2). The island-wide median output value was 0.0016 ± 0.1 (\pm SD) and the island-wide average was 0.04 ± 0.1 (range across the island: 0 - 0.94; Figure 1, Figure 2A). Low, intermediate, and highly suitable habitat is restricted to small areas throughout the island (Low [i.e., 0.2-0.4]: 4.4% of the island; Intermediate [0.4-0.6]: 1.5%; High [>0.6]: 1.0%). Highly suitable habitat patches were relatively small and aggregated (Mean Patch Size $=$ 1.4 \pm 3.3 ha; Table 1). The 10 \times 10 m locations where salamanders were detected (*n* = 476) were in highly suitable habitat (median \pm SD: 0.60 \pm 0.24, range = 0.02-0.85; ~383x higher than island-wide average). By comparison, randomly-selected locations across the island ($n = 476$) were largely in unsuitable habitat (0.0017 \pm 0.1, range $= 0 - 0.93$), indicating that our model captured relevant spatial patterns in salamander habitat suitability. These results were further corroborated by our salamander detections under coverboards: of 106 boards placed in what was deemed suitable habitat, salamanders were detected under 21.7% of these boards, whereas detections were only obtained under 3.9% of boards that were deliberately placed in apparently poor habitat (forested habitat but far from breeding locations; salamanders were only detected under 2 of these 51 boards). Finally, the Maxent model also identified that breeding locations (*n* = 33) were

comprised of highly suitable habitat (0.75 ± 0.13 , range: $0.05 - 0.88$; 479x higher than island-wide average).

Distance from breeding location was the strongest predictor of habitat suitability, contributing 59.9% to Maxent model fit, followed by land cover type (29.2%), soil type (8.4%) and elevation (2.5%) (Table 2; Figure S7). Habitat suitability decreased dramatically with distance from a breeding location, with an ~85% decline in predicted suitability upon reaching ~ 2.1 km from the center of the breeding location (Figure 1A). By ~1.6 km from the center of the breeding location, habitat suitability reached the island-wide average. Landcover types most strongly associated with suitable habitat were forested areas and swamps (predicted probability of suitable conditions were 41.0% and 31.7%, respectively; Figure S7), but forests and swamps were predicted to be unsuitable when they were distant (i.e., >1 km) from known breeding locations (0.004 \pm 0.01, range: 0 - 0.09). Agricultural areas and roads were the least suitable habitat (1.8% and 7.9%, respectively; Figure S7). In general, poorly drained soils and areas with moderate elevation were most suitable (Figure S7). The estimated suitability of habitat within a 300 m buffer zone around a breeding location was surprisingly low (0.14 ± 0.22) , range: 0.0005 - 0.94; 90x higher than island-level average). Even after reducing this buffer zone to 150 m around breeding locations, mean habitat suitability of these areas is lower than expected $(0.35 \pm 0.24$, range: 0.002 - 0.94; 220x higher than island-level average). That said, nearly all the highly suitable habitat on the island (Maxent value >0.6) is within the 150 m buffer around breeding locations (98.7%).

Overall Habitat Connectivity (All Ambystoma)
Salamander habitat connectivity was low across the island (median output value \pm SD [output values are measured in amperes]: 377.4 ± 326.2 ; range: 0 - 9108; Figure 2B) and current flow across a randomly-selected group of locations ($n = 476$) was only 2.8% lower (388.2 \pm 308.8; 1.03x higher than island-wide median). Terrestrial habitat surrounding breeding locations (i.e., within a 300 m buffer extending radially from wetland edges, excluding areas extending into Lake Erie) had moderately high current $(560.6 \pm 639.2, \text{range: } 47 - 9108; 1.5x \text{ higher than island-wide average})$. Spatially-filtered locations where salamanders were detected $(n = 476)$ occurred in areas with relatively high habitat connectivity (3307.3 \pm 2132.7, range: 850-9108; 8.8x higher than islandwide average). Forested and tallgrass communities had moderately high current flow (Forest = 599.3 \pm 485.2, 1.6x higher than island-wide average; Tallgrass = 440.0 \pm 263.2, 1.2x higher than island-wide average). Most of the known salamander sites are separated from one another by >600 m (mean \pm SD = 2.1 \pm 1.3 km), with the area between them being primarily agricultural land with low current flow (median output value \pm SD: 348.2 \pm 123.6; 0.9x lower than island-wide average).

The Least-Cost Paths analysis identified potential routes between core areas, with long stretches occurring in forested areas between farmland (42.5%), as well as through agricultural areas (22.5%) (median path length \pm SD = 4564.7 \pm 2948.7 m). Paths between breeding locations within the same core area (which were the only pathways <1 km), had high current flow (median output value \pm SD [output values are measured in amperes] $= 1092.7 \pm 1096.0$; 2.9x higher than initial habitat connectivity model) and were primarily restricted to wetland and tallgrass habitat (39.2% and 34.6% respectively). Paths between core areas (which were pathways >1km) had moderate current flow (606.5

 \pm 496.6; 1.6x higher than initial habitat connectivity model). We also examined habitat suitability of potential dispersal routes identified by the Least-Costs Path analysis by examining their habitat suitability (i.e., Maxent output values). While these routes appeared to be composed of intermediately suitable habitat (median habitat suitability \pm SD: 0.33 ± 0.19 , range: $0.0 - 0.87$; median is $210x$ higher than island-level average; Figure 2B), most of the area these routes traversed was unsuitable habitat (Unsuitable [i.e., <0.2]: 81.6% of the total area of paths; Low [0.2-0.4]: 5.6%; Intermediate [0.4-0.6]: 8.2%; High [>0.6]: 4.6%). Routes between breeding locations within the same core area, which were exclusively comprised of pathways <1 km in length, had higher habitat suitability (median \pm SD: 0.58 \pm 0.24, range: 0.05-0.86; 366.4x higher than island-level average). Paths <1 km primarily traversed intermediately and highly suitable habitat (Unsuitable [i.e., <0.2]: 13.2% of the total area of paths; Low [0.2-0.4]: 1.6%; Intermediate [0.4-0.6]: 47.9%; High [>0.6]: 37.2%).

Habitat Suitability and Connectivity across Genomotypes

We found important differences in predicted habitat suitability across genomotypes (Figure 3), even though habitat suitability of all adult salamanders was mostly governed by the same variables. Distance from breeding location was the best single predictor for all genomotypes, although this variable was substantially more important for *A. texanum* than diploid unisexuals (Figure 1B; TT: 86.4%, LT: 54.9%, LTT: 71.8%; Table 2). Habitat suitability decreased markedly, particularly for *A. texanum*, based on distance from breeding location. Specifically, habitat suitability reaches the island-wide average for *A. texanum* at 0.4 km, LT unisexuals at 1.9 km, and at 1.5 km for LTT unisexuals (Figure 1B). Highly suitable habitat (>0.6) patches were

largest for *A. texanum* but were farthest apart, less numerous and less dense compared to patches for unisexuals (see Table 1). Habitat patches for diploid unisexuals were, on average, smallest but were much closer than patches for triploid unisexuals and *A. texanum*, and were denser and more numerous (see Table 1). The amount of highly suitable habitat available in the landscape was greatest for diploid unisexuals and least for *A. texanum* (see Table 1). The second-best predictor for *A. texanum* was soil type (TT: 10.2%, LT: 7.9%, LTT: 8.1%; Table 2), with marsh soil type being identified as the most suitable (Figure S9). In contrast, land cover was the next best predictor for both diploid and triploid unisexual types (TT: 3.4%, LT: 35.1%, LTT: 18.1%; Table 2), with forest cover type being the most suitable for each group (TT: 71.3%, LT: 52.3%, LTT: 42.8%; Table 2). Highly suitable habitat (>0.6) was most isolated for *A. texanum* compared to unisexuals, with highly suitable habitat patches for LT unisexuals being more isolated than LTT unisexuals (Figure 4; Mean Proximity Index: $TT = 18.2$, $LT = 29.8$, LTT = 46.6; Mean Nearest Neighbour: TT = 1336.1 m, LT = 84.5 m, LTT = 342.3 m; MPS: TT $= 4.1$ ha, LT = 1.7 ha, LTT = 2.5 ha; Table 1).

Habitat Suitability and Connectivity across Life Stage and Seasonality

To assess the sensitivity of our observations to salamander life stage and seasonal differences, we generated separate models for adults $(n = 215)$ and juveniles $(n = 169)$. These models revealed that for both groups, distance to breeding location was the primary predictor (Figure S7; percent contribution to model: Adult $= 54.4\%$, Juveniles $=$ 81.5%, Table 2). Adults occurred in intermediately-suitable to highly-suitable habitat $(0.57 \pm 0.24$, range: 0.03-0.88), but juveniles were largely restricted to highly-suitable habitat in proximity to breeding locations $(0.68 \pm 0.26$, range = 0.006-0.89). Soil type

was more influential than land cover type for both groups (Adult soil: 20.3%; Adult land cover: 18.8%; Juvenile soil: 10.0%; Juvenile land cover 8.1%). Forests and wetlands were primary habitat features for adults (Forest: 48.9%, Wetland: 41.5%; Figure S10), but while forests remained important for juveniles, wetland importance was surprisingly lower (Forest: 49.9%, Wetland: 29.6%; Figure S10). Although salamander habitat suitability spatially varied seasonally, distance to breeding location was consistently the primary predictor of suitability (Figure S8; Mar-Apr: 60.9%, May-Jul: 68.1%, Aug-Sep: 71.2%, Table 2) and forest landcover was the primary suitable habitat type (Mar-Apr: 33.3%, May-Jul: 45.4%, Aug-Sep: 29.8%; Figure S7); wetlands were an especially important landcover type in late summer (Mar-Apr: 19.3%, May-Jul: 31.8%, Aug-Sep: 37.2%; Figure S7).

Discussion:

We evaluated suitability and connectivity of *Ambystoma* habitat on Pelee Island and found that the amount of suitable terrestrial habitat currently available for salamanders is small and mostly adjacent to breeding locations. Forested areas and nonbreeding wetlands are only important when contiguous with breeding locations or within <1 km. Habitat suitability varied across salamander genomotypes, with *A. texanum* having the least available habitat, being most associated with breeding locations and other wetlands, and also having lower habitat connectivity compared to the unisexual *Ambystoma*. LTT triploid unisexuals have a narrower habitat niche than their diploid (LT) counterparts, whereas habitat suitability for both adults and juveniles of all groups becomes less tied to wetlands as the summer progresses. Connectivity among remaining patches of suitable habitat on the island is low, and inter-patch corridors are both

prohibitively long and poor-quality to maintain functional connectivity. Overall, our findings emphasize the need for habitat protection and enhancement (primarily wetland breeding locations and nearby forest) if the long-term viability of this unique salamander complex is to remain a priority. More broadly, our analysis reveals important challenges facing salamanders when they require connectivity across human-dominated landscapes.

Many of the salamander breeding locations on Pelee Island are adjacent to, or surrounded by, unsuitable habitat (e.g., agricultural land), which has created distinct islands of suitable habitat with low connectivity to other suitable areas. Despite this, the occupied areas mostly contain a diversity of salamander genomotypes, including parental species (mainly *A. texanum*, occasionally *A. laterale*) and various types of unisexual *Ambystoma*. While additional forested habitat is available on the island, breeding locations are apparently absent from much of these areas and consequently, they do not appear to function as habitat. Indeed, the biphasic life history of most *Ambystoma* salamanders requires both suitable aquatic and terrestrial habitat, which imposes constraints on their habitat availability and suitability. Further, it is recognized that amphibian species, including *A. texanum*, avoid disturbed landscapes like agricultural fields and juveniles may migrate through areas outside the area of primary habitat suitability after emerging from their natal site (Walston and Mullin 2008). Interestingly, Greenwald et al. (2009) found that agricultural land contributed to *Ambystoma* population isolation, reinforcing the challenges faced by salamanders inhabiting human-dominated landscapes.

Paths between salamander breeding locations and surrounding areas were longer than could realistically be traversed by *A. texanum* or unisexual *Ambystoma* (Williams

1973; Denton et al. 2017), implying that there is limited movement among core sites. Most terrestrial *Ambystoma* salamanders can travel ~160 m in a single movement session (Williams 1973; Denton et al. 2017), which suggests that functional connectivity between sites requires suitable habitat at regular intervals. This does not appear to be the case across much of Pelee Island, and metapopulation dynamics may be limited or absent for these *Ambystoma* salamanders. Several of the identified paths between breeding locations traverse hedgerows between agricultural fields and these may be too long and narrow to function as effective habitat corridors for salamanders, particularly for *A. texanum* (e.g., see Vos et al. 2002). We acknowledge that evaluating the maximum useable distance of corridors based on species dispersal capabilities does not constitute a stand-alone connectivity analysis (e.g., see Baguette et al. 2013; Cushman et al. 2013), but our approach provides a crucial first step in understanding population connectivity for Pelee Island *Ambystoma* (e.g., Sawyer et al. 2011).

The observed differences between suitable habitat and connectivity for *A. texanum* and unisexuals emphasize the need for forested wetlands close to breeding locations. Patterns of suitable habitat for *A. texanum* are consistent with previous reports that this species tends to remain particularly close to their breeding sites (Williams 1973). Although some identified habitat patches for *A. texanum* were large and could be considered less fragmented, they are more isolated than patches for unisexuals, and interpatch movement is unlikely. Importantly, we found that only three of 10 core sites across Pelee Island were highly suitable for *A. texanum* species, and we know of 1-2 other breeding locations that may contain *A. laterale*, the other parental species on the island (COSEWIC 2017). In contrast, the habitat at 7 of the sites where we found salamander

communities was deemed unsuitable for *A. texanum*, although *A. texanum* was still detected in low abundance at these sites (Bare et al., in review). Breeding sites within these 7 sites also continue to produce larvae so we can infer the presence of male bisexual *Ambystoma*, though likely at a low abundance (G. Smith, unpublished data). These sites are at risk of extirpation if low habitat suitability and low connectivity to source populations compromise the viability of the host species population (see Bogart et al. 2017; Bogart 2019b). The restricted habitat requirements of *A. texanum* compared to other members of the salamander complex on Pelee Island (see also Vrijenhoek 1989; Greenwald et al. 2016) translate to a smaller total amount of suitable habitat and lower connectivity between suitable habitat patches. This presents a challenge in maintaining salamander diversity at most sites on the island where *A. texanum* is the only host available. Likewise, the variability in salamander habitat suitability across age classes and seasons indicates that habitat suitability is dynamic through time and this variability should be considered explicitly in conservation efforts.

The identification of suitable habitat and potential corridors in this study provides critical information necessary for identifying strategies and locations for habitat restoration necessary for the persistence of *Ambystoma* salamanders on Pelee Island. Their long-term viability may require conservation strategies similar to those recommended by Semlitsch & Bodie (2003) and Calhoun et al. (2005), which include the protection and enhancement of suitable habitat near current breeding locations. Scott et al. (2013) also emphasize the importance of terrestrial buffers around breeding locations. Recent work on endangered *A. jeffersonianum* advocates for protection of 400-450 m of terrestrial habitat around breeding locations (Van Drunen et al. 2020). Similarly, we find

that most of the suitable habitat for *Ambystoma* salamanders on Pelee Island, as well as most of the individual salamanders, are found within 450 m of a breeding site and protecting a similar amount of habitat around breeding locations would assist the longterm persistence of *A. texanum* and unisexual *Ambystoma*.

Given *A. texanum*'s role as the primary reproductive host in this system and their limited suitable habitat we now consider the possible management implications. To increase the amount of suitable habitat, establishing breeding locations within 1 km of occupied sites, but still away from unusable habitat, should bolster population viability and the persistence of this complex. However, constructed ponds may remain unsuitable for 15-20+ years (Mitsch and Wilson 1996), so site selection and construction of new breeding locations must be strategic and accompanied by additional conservation management efforts (Ward and Hossie 2020). On Pelee Island, re-establishing functional connectivity among patches of suitable habitat remains a major challenge because most land outside core breeding locations and adjacent forest is on private land, meaning that incentives may be necessary to encourage natural landcover restoration and conservation. Regardless, it is important to note that given the current scale of salamander habitat fragmentation on Pelee Island, it may simply not be possible to restore functional metapopulation dynamics. More drastic interventions like head-starting juveniles and assisted dispersal may need to be considered as a means to maintain the viability of the host populations which support this complex (Seddon et al. 2014; O'Donnell et al. 2017). Accordingly, over the longer term it may be more effective to bolster the protection of suitable salamander habitat within currently-occupied sites, which is consistent with the outcome of other habitat suitability and connectivity modelling studies in humandominated landscapes (e.g., see Compton et al. 2007; Pereira et al. 2011; Crooks et al. 2011; Rondinini et al. 2011).

Ultimately, these findings showcase the importance of identifying suitable habitat for species existing in a fragmented system. Future studies on this *Ambystoma* complex should focus on environmental monitoring to evaluate the effectiveness of *in situ* conservation efforts, such as the construction of new breeding locations and reforestation. We concur with the recommendations of De Kort et al. (2020), in that the accuracy of habitat suitability models improves when accounting for species dispersal, as well as by validating model results in unsampled areas. Our results also suggest that a targeted focus on the habitat requirements for key members of a community or assemblage in conservation and restoration planning can be beneficial, as their protection can function as a conservation umbrella and protect other species (Johnson et al. 2017). Indeed, given the rapid decline in biodiversity globally (Hoffmann et al. 2010), ecosystem and community-based approaches to identifying and protecting key habitat are increasingly needed to inform and guide conservation efforts.

Tables and Figures

Table 1: Patch-level, class-level and landscape-level metrics measured with Fragstats for highly suitable habitat for *Ambystoma texanum* and unisexual Ambystoma on Pelee Island, ON. All metrics were measured at the class-level, aside from AREA and PERIM measured at patch-level and GYRATE measured at landscape-level. See main text for metric descriptions. The model with all locations was created with data from 2015-2019, and genomotype-specific models were created with data from 2015-2018.

Table 2: Percent contribution and permutation importance of selected environmental variables in Maxent models for *A. texanum* and unisexual *Ambystoma* captured from 2015 – 2019, and from 2015-2018 for genomotype-specific models.

Model	Variable	Percent	Permutation
		Contribution	Importance
All Locations	Distance to Pond	59.9	72.8
	Land Cover Type	29.2	18.1
	Soil Type	8.4	5.4
	Elevation	2.5	3.7
Adults	Distance to Pond	54.4	64.3
	Land Cover Type	18.8	18.9
	Soil Type	20.3	12.9
	Elevation	6.5	3.9
Juveniles	Distance to Pond	81.5	89.3
	Land Cover Type	8.1	6.2
	Soil Type	10.0	3.7
	Elevation	0.5	0.8
Ambystoma texanum	Distance to Pond	86.4	98.4
(TT)	Land Cover Type	3.4	0.0
	Soil Type	10.2	1.6
	Elevation	0.0	0.0
Unisexual	Distance to Pond	54.9	66.5
Ambystoma (LT)	Land Cover Type	35.1	23.6
	Soil Type	7.9	6.9
	Elevation	2.1	3.0
Unisexual	Distance to Pond	71.8	87.8
Ambystoma (LTT)	Land Cover Type	18.1	5.1
	Soil Type	8.1	5.3
	Elevation	2.1	1.8
March - April	Distance to Pond	60.9	66.7
captures	Land Cover Type	29.8	20.9
	Soil Type	6.7	8.9
	Elevation	2.5	3.5
$May - July captures$	Distance to Pond	68.1	77.2
	Land Cover Type	14.2	11.4
	Soil Type	16.1	9.9
	Elevation	1.6	1.4
August - September	Distance to Pond	71.2	78.7
captures	Land Cover Type	15.5	10.2
	Soil Type	13.0	10.5
	Elevation	0.3	0.5

Figure 1: Relationship between distance to breeding location (m) and predicted habitat suitability for *Ambystoma* salamanders on Pelee Island, ON, Canada (based on response curves from Maxent models). A) The relationship between distance to breeding site and predicted habitat suitability based on spatially filtered location data ($n = 476$) for all adults and juvenile salamanders collected from 2015-2019. B) The relationship between distance to breeding site and predicted habitat suitability for three distinct *Ambystoma* genomotypes, from data collected from 2015-2018. Dotted line depicts the relationship for diploid (LT) unisexual *Ambystoma* salamanders, the dashed line depicts the relationship for triploid (LTT) unisexual *Ambystoma* salamanders, and the solid line depicts the relationship for *A. texanum* (TT) salamanders. Higher values on y-axis represent higher probability of presence.

Figure 2: A) Predicted habitat suitability for *Ambystoma* salamanders on Pelee Island, ON, Canada using spatially filtered location data ($n = 476$) collected from 2015-2019 (as modelled in Maxent). Environmental predictor variables include distance to breeding site, land cover type, soil type, and elevation. Green indicates areas with predicted high habitat suitability and red indicates areas with low habitat suitability. B) Predicted habitat connectivity for *Ambystoma* salamanders on Pelee Island, ON, Canada produced using spatially filtered localities $(n = 476)$ and a land cover raster grid using resistance values based on Compton et al. 2007 (see Table S1). Green indicates well-connected areas and red indicate areas with low connectivity. Lines depict predicted least-cost paths among known communities, with paths under 1 km in length depicted in purple, and paths are over 1 km in black.

Figure 3: Predicted habitat suitability for three *Ambsytoma* salamander genomotypes on Pelee Island, ON, Canada, using spatially filtered location data (from 2015-2018) and environmental predictor variables including distance to breeding site, land cover type, soil type, and elevation. Left panel: Predicted habitat suitability for *Ambystoma texanum* (TT) salamanders. Middle panel: Predicted habitat suitability for diploid (LT) unisexual *Ambystoma* salamanders. Right panel: Predicted habitat suitability for triploid (LTT) unisexual *Ambystoma* salamanders. Green indicates areas with predicted high habitat suitability, with orange and red indicating areas with low habitat suitability.

Figure 4: Predicted habitat connectivity for three *Ambystoma* salamander genomotypes on Pelee Island, ON, Canada. The models used spatially filtered localities (from 2015-2018) and a land cover raster grid with resistance values based on Compton et al. 2007 (see Table S1). Left panel: Connectivity for *A. texanum* (TT) salamanders. Middle panel: Connectivity for diploid (LT) unisexual *Ambystoma* salamanders. Right panel: Connectivity for triploid (LTT) unisexual *Ambystoma* salamanders. Green indicates well-connected areas, with orange and red indicating areas with low connectivity.

Acknowledgements:

The Government of Ontario provided financial support for this work through the SARSF and SARRFO programs (SAR-00109, SARSF_23_18_Dmurr2, RF_23_18_Trent6). Nature Conservancy of Canada, Essex Region Conservation Authority, Ontario Nature, Ontario Parks, as well as J. Ambrose, J. DeMarco, and D. Kraus, provided authorization to sample on their properties. We thank P. Heney, A. Myette, J. Leavitt, C. Watt, M. Ward and Scales Nature Park for assistance with field work. Animals were handled in accordance with Animal Care protocols approved by Trent University and Ontario Ministry of Natural Resources (Protocols: 23906, 25301, 25344, 25670). Land access permits, Wildlife Scientific Collector's Authorizations (1079527, 1082275, 1085623, 1088782, 1092367), and a Notice of Activity under the Ontario Endangered Species Act (Confirmation ID: M-102-3802796883, M-102-9225853169, M-102-1254262277) were secured to conduct this work.

References

- Atmoko T, Mardiastuti A, Bismark M, et al (2020) Habitat suitability of Proboscis monkey (Nasalis larvatus) in Berau Delta, East Kalimantan, Indonesia. Biodiversitas 21:5155–5163. https://doi.org/10.13057/biodiv/d211121
- Baguette M, Blanchet S, Legrand D, et al (2013) Individual dispersal, landscape connectivity and ecological networks. Biol Rev 88:310–326. https://doi.org/10.1111/brv.12000
- Becker CG, Fonseca CR, Haddad CFB, et al (2007) Habitat Split and the Global Decline of Amphibians. Science 318:1775–1777. https://doi.org/10.1126/science.1149374
- Beier P, Noss RF (1998) Do Habitat Corridors Provide Connectivity? Conserv Biol 12:1241–1252
- Bennett AF (2003) Linkages in the Landscape: The Role of Corridors and Connectivity in Wildlife Conservation, 2nd edn. IUCN, Gland, Switzerland and Cambridge, UK
- Bogart J, Bi K, Fu J, et al (2007) Unisexual salamanders (genus Ambystoma) present a new reproductive mode for eukaryotes. Genome 50:119–136. https://doi.org/10.1139/G06-152
- Bogart J, Licht L, Oldham M, Darbyshire S (1985) Electrophoretic identification of Ambystoma laterale and Ambystoma texanum as well as their diploid and triploid interspecific hybrids (Amphibia: Caudata) on Pelee Island, Ontario. Can J Zool 63:340–347. https://doi.org/10.1139/z85-052
- Bogart J, Licht LE (1986) Reproduction and the origin of polyploids in hybrid salamanders of the genus *Ambystoma*. Can J Genet Cytol 28:605–617. https://doi.org/10.1139/g86-089
- Bogart J, Linton J, Sandilands A (2017) A Population in Limbo: Unisexual Salamanders (Genus Ambystoma) Decline Without Sperm-donating Species. Herpetol Conserv Biol 12:41–55
- Bogart JP (2019a) A family study to examine clonal diversity in unisexual salamanders (genus Ambystoma). Genome 62:549–561. https://doi.org/10.1139/gen-2019- 0034
- Bogart JP (2019b) Unisexual Salamanders in the Genus Ambystoma. Herpetologica 75:259. https://doi.org/10.1655/Herpetologica-D-19-00043.1
- Bogart JP, Bartoszek J, Noble DWA, Bi K (2009) Sex in unisexual salamanders: discovery of a new sperm donor with ancient affinities. Heredity 103:483–493. https://doi.org/10.1038/hdy.2009.83
- Callaway RM (2007) Positive Interactions and Interdependence in Plant Communities. Springer Science & Business Media, Dordrecht, Netherlands
- Calhoun AJK, Miller NA, Klemens MW (2005) Conserving pool-breeding amphibians in human-dominated landscapes through local implementation of Best Development

Practices. Wetlands Ecol Manage 13:291–304. https://doi.org/10.1007/s11273- 004-7523-8

- Compton BW, McGarigal K, Cushman SA, Gamble LR (2007) A Resistant-Kernel Model of Connectivity for Amphibians that Breed in Vernal Pools. Conserv Biol 21:788–799. https://doi.org/10.1111/j.1523-1739.2007.00674.x
- COSEWIC (2017) Unisexual ambystoma (Ambystoma laterale) and 3 subspecies: COSEWIC assessment and status report 2016. In: Gov. Can. https://www.canada.ca/en/environment-climate-change/services/species-riskpublic-registry/cosewic-assessments-status-reports/unisexual-ambystomasubspecies-2016.html. Accessed 6 Jan 2021
- Crooks KR, Burdett CL, Theobald DM, et al (2011) Global patterns of fragmentation and connectivity of mammalian carnivore habitat. Philos Trans R Soc B Biol Sci 366:2642–2651. https://doi.org/10.1098/rstb.2011.0120
- Crooks KR, Sanjayan M (eds) (2006) Connectivity Conservation. Cambridge University Press, Cambridge UK
- Cushman SA, McRae B, Adriaensen F, et al (2013) Biological corridors and connectivity. In: Macdonald DW, Willis KJ (eds) Key Topics in Conservation Biology 2. John Wiley & Sons, Oxford, pp 384–404
- Davidson NC (2014) How much wetland has the world lost? Long-term and recent trends in global wetland area. Mar Freshw Res 65:934. https://doi.org/10.1071/MF14173
- De Kort H, Baguette M, Lenoir J, Stevens VM (2020) Toward reliable habitat suitability and accessibility models in an era of multiple environmental stressors. Ecol Evol 10:10937–10952. https://doi.org/10.1002/ece3.6753
- Denton RD, Greenwald KR, Gibbs HL (2017) Locomotor endurance predicts differences in realized dispersal between sympatric sexual and unisexual salamanders. Funct Ecol 31:915–926. https://doi.org/10.1111/1365-2435.12813
- Dixon MJR, Loh J, Davidson NC, et al (2016) Tracking global change in ecosystem area: The Wetland Extent Trends index. Biol Conserv 193:27–35. https://doi.org/10.1016/j.biocon.2015.10.023
- Dondina O, Saura S, Bani L, Mateo-Sánchez MC (2018) Enhancing connectivity in agroecosystems: focus on the best existing corridors or on new pathways? Landsc Ecol 33:1741–1756. https://doi.org/10.1007/s10980-018-0698-9
- Ducks Unlimited Canada (2010) Southern Ontario Wetland Conversion Analysis: Final Report. Barrie, ON
- Elith J, Phillips SJ, Hastie T, et al (2011) A statistical explanation of MaxEnt for ecologists. Divers Distrib 17:43–57. https://doi.org/10.1111/j.1472- 4642.2010.00725.x
- Elkie PC, Rempel RS, Carr AP (1999) Patch Analyst User's Manual: A tool for quantifying landscape structure. Northwest Science and Technology, Thunder Bay, ON
- Environment and Climate Change Canada (2020) Recovery strategy for the smallmouthed salamander (Ambystoma texanum) in Canada. Environment and Climate Change Canada, Ottawa, ON
- Environmental Commissioner of Ontario (2018) Back to the basics: Southern Ontario's wetlands and forests. Government of Ontario, Toronto, ON
- Fischer J, Lindenmayer DB (2007) Landscape modification and habitat fragmentation: a synthesis. Glob Ecol Biogeogr 16:265–280. https://doi.org/10.1111/j.1466- 8238.2007.00287.x
- Gallo JA, Greene R (2018) Connectivity Analysis Software for Estimating Linkage Priority. https://doi.org/10.6084/m9.figshare.5673715.v5
- Gamble LR, McGarigal K, Compton BW (2007) Fidelity and dispersal in the pondbreeding amphibian, Ambystoma opacum: Implications for spatio-temporal population dynamics and conservation. Biol Conserv 139:247–257. https://doi.org/10.1016/j.biocon.2007.07.001
- Gopurenko D, Williams RN, DeWoody JA (2007) Reproductive and Mating Success in the Small-Mouthed Salamander (Ambystoma texanum) Estimated via Microsatellite Parentage Analysis. Evol Biol 34:130–139. https://doi.org/10.1007/s11692-007-9009-0
- Greenwald KR, Denton RD, Gibbs HL (2016) Niche partitioning among sexual and unisexual Ambystoma salamanders. Ecosphere 7:e01579. https://doi.org/10.1002/ecs2.1579
- Greenwald KR, Gibbs HL, Waite TA (2009) Efficacy of Land-Cover Models in Predicting Isolation of Marbled Salamander Populations in a Fragmented Landscape. Conserv Biol 23:1232–1241. https://doi.org/10.1111/j.1523- 1739.2009.01204.x
- Haddad NM, Brudvig LA, Clobert J, et al (2015) Habitat fragmentation and its lasting impact on Earth's ecosystems. Sci Adv 1:e1500052. https://doi.org/10.1126/sciadv.1500052
- Hanski I (1998) Metapopulation dynamics. Nature 396:41–49. https://doi.org/10.1038/23876
- Harding JH, Mifsud DA (2017) Amphibians and Reptiles of the Great Lakes Region, Revised Ed. University of Michigan Press, Ann Arbor, Michigan
- Hatcher MJ, Dick JT, Dunn AM (2012) Diverse effects of parasites in ecosystems: linking interdependent processes. Front Ecol Environ 10:186–194. https://doi.org/10.1890/110016
- Hecnar SJ (2004) Great Lakes wetlands as amphibian habitats: A review. Aquat Ecosyst Health Manag 7:289–303. https://doi.org/10.1080/14634980490461542
- Hoffmann M, Hilton-Taylor C, Angulo A, et al (2010) The Impact of Conservation on the Status of the World's Vertebrates. Science 330:1503–1509. https://doi.org/10.1126/science.1194442
- Hossie TJ (2018) Recovery Strategy for Small-mouthed Salamander (Ambystoma texanum) and Unisexual Ambystoma Small-mouthed Salamander dependent population (Ambystoma laterale - texanum) in Ontario. Ontario Ministry of Natural Resources and Forestry, Peterborough, Ontario.
- Johnson SA, Ober HK, Adams DC (2017) Are keystone species effective umbrellas for habitat conservation? A spatially explicit approach. J Nat Conserv 37:47–55. https://doi.org/10.1016/j.jnc.2017.03.003
- Jutterbock JE, Owen PC (2013) Blue-spotted salamander, Ambystoma laterale (Hallowell 1856). In: Pfingsten RA, Davis JG, Matson TO, et al. (eds) Amphibians of Ohio. Ohio Biological Survey, Columbus, Ohio, p 899
- Keymer JE, Marquet PA, Velasco‐Hernández JX, et al (2000) Extinction Thresholds and Metapopulation Persistence in Dynamic Landscapes. The American Naturalist 156:478–494. https://doi.org/10.1086/303407
- Kolozsvary MB, Swihart RK (1999) Habitat fragmentation and the distribution of amphibians: patch and landscape correlates in farmland. Can J Zool 77:1288– 1299
- Lehtinen RM, Galatowitsch SM, Tester JR (1999) Consequences of habitat loss and fragmentation for wetland amphibian assemblages. Wetlands 19:1–12. https://doi.org/10.1007/BF03161728
- Lowcock LA, Licht LE, Bogart JP (1987) Nomenclature in Hybrid Complexes of Ambystoma (Urodela: Ambystomatidae): No Case for the Erection of Hybrid "Species." Syst Zool 36:328–336. https://doi.org/10.2307/2413070
- Madison DM (1997) The Emigration of Radio-Implanted Spotted Salamanders, Ambystoma maculatum. J Herpetol 31:542–551. https://doi.org/10.2307/1565607
- Mantyka‐Pringle CS, Martin TG, Rhodes JR (2012) Interactions between climate and habitat loss effects on biodiversity: a systematic review and meta-analysis. Glob Change Biol 18:1239–1252. https://doi.org/10.1111/j.1365-2486.2011.02593.x
- Marsh DM, Trenham PC (2001) Metapopulation Dynamics and Amphibian Conservation. Conserv Biol 15:10
- McGarigal K (2015) FRAGSTATS help. https://ibis.geog.ubc.ca/courses/geob479/labs/fragstats.help.4.pdf. Accessed 17 Sep 2021
- McRae BH, Dickson BG, Keitt TH, Shah VB (2008) Using circuit theory to model connectivity in ecology, evolution, and conservation. Ecology 89:2712–2724. https://doi.org/10.1890/07-1861.1
- McRae BH, Kavanagh DM (2011) Linkage Mapper Connectivity Analysis Software. http://www.circuitscape.org/linkagemapper. Accessed 16 Jan 2021
- McRae BH, Shah V, Mohapatra T (2014) Circuitscape User Guide. http://docs.circuitscape.org/circuitscape_4_0_user_guide.html?&id=gsite. Accessed 16 Jan 2021
- Mills PB, Hossie TJ, Murray DL (2020) Niche determinants in a salamander complex: Does hybridism or reproductive parasitism explain patterns of distribution? Ecosphere 11:e03265. https://doi.org/10.1002/ecs2.3265
- Mitsch W, Wilson R (1996) Improving the Success of Wetland Creation and Restoration with Know-How, Time, and Self-Design. Ecol Appl 6:77. https://doi.org/10.2307/2269554
- O'Donnell KM, Messerman AF, Barichivich WJ, et al (2017) Structured decision making as a conservation tool for recovery planning of two endangered salamanders. J Nat Conserv 37:66–72. https://doi.org/10.1016/j.jnc.2017.02.011
- Ontario Ministry of Agriculture, Food, and Rural Affairs (OMAFRA) (2019) Land Information Ontario Data Description. Ontario Ministry of Agriculture, Food, and Rural Affairs, Guelph, ON
- Owen PC, Jutterbock JE (2013) Small-mouthed salamander, Ambystoma texanum (Mathes 1855). In: Pfingsten RA, Davis JG, Matson TO, et al. (eds) Amphibians of Ohio. Ohio Biological Survey, Columbus, Ohio, p 899
- Pereira M, Segurado P, Neves N (2011) Using spatial network structure in landscape management and planning: A case study with pond turtles. Landsc Urban Plan 100:67–76. https://doi.org/10.1016/j.landurbplan.2010.11.009
- Peterman WE, Connette GM, Spatola BN, et al (2012) Identification of Polymorphic Loci in Ambystoma annulatum and Review of Cross-species Microsatellite Use in the Genus Ambystoma. Copeia 2012:570–577. https://doi.org/10.1643/CH-11-001
- Petranka JW (1998) Salamanders of the United States and Canada. Smithsonian Institution Press, Washington and London
- Phillips SJ (2017) A Brief Tutorial on Maxent. https://biodiversityinformatics.amnh.org/open_source/maxent/Maxent_tutorial_20 21.pdf. Accessed May 5 2021
- Pierik ME, Dell'Acqua M, Confalonieri R, et al (2016) Designing ecological corridors in a fragmented landscape: A fuzzy approach to circuit connectivity analysis. Ecol Indic 67:807–820. https://doi.org/10.1016/j.ecolind.2016.03.032
- Porej D, Hetherington TE (2005) Designing Wetlands for Amphibians: The Importance of Predatory Fish and Shallow Littoral Zones in Structuring of Amphibian Communities. Wetl Ecol Manag 13:445–455. https://doi.org/10.1007/s11273-004- 0522-y
- Porej D, Micacchion M, Hetherington TE (2004) Core terrestrial habitat for conservation of local populations of salamanders and wood frogs in agricultural landscapes. Biol Conserv 120:399–409. https://doi.org/10.1016/j.biocon.2004.03.015
- Provincial Mapping Unit (2018) Provincial Digital Elevation Model (PDEM) User Guide.

https://geohub.lio.gov.on.ca/datasets/882a9059ec7c4881abbdb6afa0ae73e6. Accessed 5 May 2021

- Rondinini C, Di Marco M, Chiozza F, et al (2011) Global habitat suitability models of terrestrial mammals. Philos Trans R Soc B Biol Sci 366:2633–2641. https://doi.org/10.1098/rstb.2011.0113
- Rudnick DA, Ryan SJ, Beier P, et al (2012) The role of landscape connectivity in planning and implementing conservation and restoration priorities. Ecological Society of America, Rocky Mountain Research Station, Fort Collins, CO
- Sawyer SC, Epps CW, Brashares JS (2011) Placing linkages among fragmented habitats: do least-cost models reflect how animals use landscapes?: Least-cost modelling for habitat linkage design. J Appl Ecol 48:668–678. https://doi.org/10.1111/j.1365-2664.2011.01970.x
- Scott DE, Komoroski MJ, Croshaw DA, Dixon PM (2013) Terrestrial distribution of pond-breeding salamanders around an isolated wetland. Ecology 94:2537–2546. https://doi.org/10.1890/12-1999.1
- Seddon PJ, Griffiths CJ, Soorae PS, Armstrong DP (2014) Reversing defaunation: Restoring species in a changing world. Science 345:406–412. https://doi.org/10.1126/science.1251818
- Semlitsch RD, Bodie JR (2003) Biological Criteria for Buffer Zones around Wetlands and Riparian Habitats for Amphibians and Reptiles. Conserv Biol 17:1219–1228. https://doi.org/10.1046/j.1523-1739.2003.02177.x
- Singh M, Tokola T, Hou Z, Notarnicola C (2017) Remote sensing-based landscape indicators for the evaluation of threatened-bird habitats in a tropical forest. Ecol Evol 7:4552–4567. https://doi.org/10.1002/ece3.2970
- SOLRIS Technical Team (2015) Southern Ontario Land Resource Information System (SOLRIS) Version 2.0: Data Specifications. Ministry of Natural Resources and Forestry, Sault Ste Marie, ON
- Thomas CD, Cameron A, Green RE, et al (2004) Extinction risk from climate change. Nature 427:145–148. https://doi.org/10.1038/nature02121
- Van Drunen SG, Linton JE, Bogart JP, et al (2020) Estimating critical habitat based on year-round movements of the endangered Jefferson Salamander (Ambystoma jeffersonianum) and their unisexual dependents. Can J Zool 98:117–126
- Vos CC, Baveco H, Grashof-Bokdam CJ (2002) Corridors and Species Dispersal. In: Gutzwiller KJ (ed) Applying Landscape Ecology in Biological Conservation. Springer New York, New York, NY, pp 84–104
- Vrijenhoek RC (1989) Genetic and Ecological Constraints on the Origins and Establishment of Unisexual Vertebrates. In: Dawley RM, Bogart JP (eds) Evolution and Ecology of Unisexual Vertebrates. New York State Museum, Albany, NY, pp 24–31
- Walston LJ, Mullin SJ (2008) Variation in amount of surrounding forest habitat influences the initial orientation of juvenile amphibians emigrating from breeding ponds. Can J Zool 86:141–146. https://doi.org/10.1139/Z07-117
- Ward M, Hossie TJ (2020) Do Existing Constructed Ponds on Pelee Island, Ontario Match the Habitat Requirements of Endangered Ambystoma Larvae? Wetlands 40:2097–2108. https://doi.org/10.1007/s13157-020-01364-8
- Williams PK (1973) Seasonal movements and population dynamics of four sympatric Mole Salamanders, genus Ambystoma. Ph.D. Thesis, Indiana University
- Williams RN, DeWoody JA (2004) Fluorescent dUTP helps characterize 10 novel tetranucleotide microsatellites from an enriched salamander (Ambystoma texanum) genomic library. Mol Ecol Notes 4:17–19. https://doi.org/10.1046/j.1471-8286.2003.00559.x
- Yuh YG, N'Goran PK, Dongmo ZN, et al (2020) Mapping suitable great ape habitat in and around the Lobéké National Park, South-East Cameroon. Ecol Evol 10:14282–14299. https://doi.org/10.1002/ece3.7027

Chapter 3: General Discussion

Thesis Goals: Revisited

This thesis aimed at identifying suitable habitat and corridors for an imperilled assemblage of salamanders, *Ambystoma* spp., across a fragmented island landscape. I determined that small amounts of suitable habitat remain on Pelee Island, and unsurprisingly, terrestrial habitat was most suitable when close to breeding locations. Habitat patches were poorly connected and most were likely too distant from one another to be realistically accessed by dispersing salamanders without habitat modification inside corridors. Of all salamander genomotypes I evaluated, habitat for *A. texanum* was most restricted and isolated whereas diploid (LT) unisexuals had the broadest habitat suitability. From a conservation perspective, increased attention will need to be paid to the status and habitat availability for *A. texanum* on Pelee Island, including through population monitoring and habitat restoration, particularly areas where corridors or suitable breeding ponds could be strategically restored or placed. Overall, our study demonstrates the value of assessing the quality of terrestrial and wetland habitat for imperilled salamanders, especially when habitat patches are small and metapopulation dynamics may be impacted by high fragmentation and limited opportunities for dispersal.

Implications for Ambystoma *Conservation Management on Pelee Island*

There is much concern over the viability of the *Ambystoma* salamander complex on Pelee Island and whether populations are sustainable over the long term (Hossie 2018). First, the persistence of this complex is especially precarious because of the interconnectedness of each group, including reliance of unisexual *Ambystoma* on the much rarer parental species for sperm parasitism. Results from this study highlight differences

in habitat preferences for different groups, meaning that protection or restoration measures must be directed collectively while also avoiding disruptively favouring one group over another. I recognize that the salamander complex requires both terrestrial and aquatic habitat, meaning that protection of areas surrounding breeding locations is essential. Currently, there is no legislation which defines protected habitat for smallmouthed salamanders or the associated unisexuals, however the provincial recovery strategy recommended protection of "all suitable terrestrial habitat extending radially 300 m from the edge of a known small-mouthed or unisexual *Ambystoma* breeding [location]" (Hossie 2018). Similarly, recent recommendations for protection of the endangered *A. jeffersonianum* salamander in southern Ontario include protecting a 400-450 m buffer around known breeding locations (Van Drunen et al. 2020). My analysis highlights the benefit of a larger buffer size, up to 400-450 m from the breeding location, to accommodate all life stages and critical behaviours like foraging or overwintering. To achieve this goal, it may be necessary to actively restore suitable habitat in protected areas in proximity to sites that are currently occupied by salamanders to bolster the small size of suitable habitat patches (i.e., wetland and forest). Semlitsch and Bodie (2003) advocate for the stratification of protection zones to include both aquatic and terrestrial buffers up to 400 m, where the terrestrial buffer extends beyond the core habitat area. This recommendation highlights the deficiency in Ontario's current land-use planning approach because wetland protection rarely encompasses accompanying terrestrial buffers of sufficient size to support adult salamanders.

Broadly speaking, current conservation and habitat restoration efforts on Pelee Island are relatively widespread, with roughly 18% of the island being protected (777.6

ha) publicly and by private landowners (including conservation easements, provincial parks, and areas managed by environmental non-governmental organizations); this compares to only 11% protection for mainland Ontario (MNRF 2017; Office of the Auditor General of Ontario 2020). The protected areas on Pelee Island include forested and wetland ecosystems like swamp forests, as well as some historical farmland which has been converted into native meadows as well as small wetlands. Some of these protected areas are relatively large, ranging in size from 9 to 183 hectares (G. Smith, unpublished data). A number of these areas are isolated, but many are well-connected and effectively serve as large conservation areas managed by multiple stakeholders. Pelee Island supports many species listed as threatened both federally and provincially, including over 100 plant species and several unique snails and herpetofauna. Pelee Island also includes several unique ecosystem types which are globally rare, including alvars and oak savannahs (Forbes et al. 2000; NCC 2008).

As a testament to the momentum for active involvement in stewardship and restoration efforts in protected areas on Pelee Island, note that the Nature Conservancy of Canada (NCC) recently constructed a \sim 25 ha wetland on their property in proximity to an existing population of salamanders. NCC constructed additional ponds in 2021 in areas identified by our team as potential movement corridors for salamanders and is also supporting natural forest succession in several open fields adjacent to existing salamander habitat (J. Crosthwaite, personal communication, 2021). My study showed that forest and non-breeding wetland ecosystems must be close to breeding locations to be considered suitable habitat for *Ambystoma* spp., so the improvement of these areas will be beneficial for the persistence of this population by increasing breeding habitat and overall

connectivity within this area. These efforts will be particularly beneficial for some of the largest populations of salamanders that I studied, as the constructed ponds and expanded forested habitat are directly adjacent. Accordingly, efforts to establish additional protected areas on Pelee Island are likely to help promote the long-term viability of the *Ambystoma* spp. complex. As evidence of this success, three newly-constructed ponds (fall 2020), which were established following recommendations informed by my preliminary research findings and modified based on follow-up recommendations, have already been used by salamanders for breeding purposes (i.e., larvae and juveniles were detected in 2021 in one breeding locations, eggs were detected in spring 2022 in all three breeding locations; J. McDonald, personal communication). This pond is >300 m from the nearest known existing *Ambystoma* spp. breeding location, providing a promising indication that this conservation effort is having tangible benefits. Future restoration efforts and research should continue to evaluate the success of these constructed ponds considering that they may not always be situated in areas that are as suitable for salamanders as natural breeding locations (Ward and Hossie 2020).

Improving HS and Connectivity Modelling Approaches

Habitat suitability and connectivity models such as those used in this thesis are predictive and assume that current conditions are stationary and that species observations used to train the model are representative. It is reasonable to assume that Pelee Island's landcover features and environmental conditions are not rapidly changing, but occurrence data are often recognized as being biased towards areas having received disproportionate survey efforts in the past; this bias can exaggerate the assessed importance of overrepresented sampling locations (Fourcade et al. 2014). It follows that such

misrepresentation can lead to severe repercussions for conservation and management (Kramer-Schadt et al. 2013). Yackulic et al. (2013) found that most (87%) of reported studies assessing species occurrence records had not applied appropriate measures to limit potential bias in their sampling methods. Notably, my work included considerable efforts to avoid similar problems by rarefying location data to minimize spatial autocorrelation of the environmental predictors used in my models. Further, because species distribution models are 'presence-only' and therefore do not use known absence data but rather generate pseudo-absences, there is a high risk of false-negative detections, especially for species that are cryptic or have seasonally variable distributions (Hirzel and Le Lay 2008; Lobo et al. 2010). However, my efforts to purposefully sample in areas assumed to be unsuitable habitat and in previously unsampled areas aimed to avoid this bias (i.e., avoid 'methodological absences'; see Lobo et al. 2010). This testing largely confirmed that areas modelled as being poor salamander habitat would not be able to support animals.

Species distribution and connectivity models are associated with a degree of uncertainty and should be interpreted based on the biology of the species of interest. Models are solely based on the input variables, and the output can be sensitive to variation in animal-environment interactions (Thuiller and Münkemüller 2010; Rudnick et al. 2012). These input variables had to be carefully selected in my analyses to only include variables relevant to *Ambystoma* spp., including avoiding the stratification of land cover types into groups that were functionally equivalent to salamanders. Connectivity models, in particular, must account for species biology, including the ability of organisms to travel a given distance in marginal habitat. It follows that field validation of this

assumption is a necessary aspect of developing corridors that are relevant to the species of interest (Rudnick et al. 2012). Particular attention should be paid to geographical barriers to movement, including unusable habitat such as roads or tilled fields (Kramer-Schadt et al. 2004; Kimmig et al. 2020). These land use changes can compound the effects of habitat fragmentation on dispersal capability, but the extent of acceptable disturbance will depend on an animal's ability to occupy and cross through marginal habitat. Given their small size and dependence on moisture for survival, *Ambystoma* salamanders may have limited ability to use marginal habitat unless it provides sufficient structural cover and protection from sun and wind.

Role of Modelling in Research and Conservation Management

Conservation planners and managers must prioritize protection and strategic integration of areas that have high ecological value, as well as identify potential corridors for maintaining metapopulation dynamics and species persistence. Habitat suitability and connectivity models can be used to evaluate the current distribution of species and determine areas for conservation priority or species reintroduction (Larson et al. 2004; Thorn et al. 2009). For *Ambystoma* spp. on Pelee Island, this could potentially involve assisted colonization or *ex situ* head-starting in intermediately- or highly-suitable areas. Such efforts could improve *Ambystoma* spp. resilience to demographic and environmental changes particularly in areas with low numbers of parental species (Semlitsch et al. 2017). Future investigation will be necessary to assess the feasibility of these recovery strategies and whether newly-established animals fully use restored habitats for reproduction or dispersal and whether viable populations can be sustained. Additionally, habitat suitability models, if designed properly, have the potential to predict areas where

unknown populations may be found; this can be particularly beneficial for focusing field surveys and conservation efforts which have finite resources (e.g., see Stratmann et al. 2016; Udyawer et al. 2020). As mentioned previously, my preliminary work identifying potentially suitable habitat along movement corridors on Pelee Island helped inform the establishment of several new *Ambystoma* breeding ponds and my final results help highlight additional candidate sites.

Conclusions and Future Directions

Broadly, my findings emphasize the importance of evaluating remaining habitat features for imperilled species occupying altered landscapes. I identified characteristics of the remaining habitats that are important for the *Ambystoma* salamander complex on Pelee Island, many of which will require increased and continued conservation management and habitat restoration planning to maintain their long-term viability. This approach would be beneficial for other species at risk of extirpation, such as the Blanding's turtle (*Emydoidea blandingii*) and the blue racer (*Coluber constrictor foxii*) on Pelee Island, or others that are threatened as a result of encroaching urban development on natural areas and have limited distributions on the island. More work needs to be done on the Pelee Island salamander complex to evaluate the success of habitat creation efforts and to monitor changes in habitat suitability and connectivity as well as population and distribution. Future investigation should also incorporate population monitoring on groups within smaller and more isolated areas of suitable habitat to assess their long-term viability. Such efforts are especially needed in populations with low abundance of parental *Ambystoma* species, where extinction risk may be high. Finally, I recommend that future studies incorporating habitat suitability and connectivity models for terrestrial

salamanders need to account for biased spatial and input data to improve the reliability of the models. These efforts will ensure the accuracy of model findings and thereby help guide effective conservation efforts in highly fragmented landscapes. In sum, my thesis makes an important contribution in this area and provides a strong foundation for future efforts targeting the conservation of salamanders and other organisms on Pelee Island and beyond.

References

- Forbes MR, Schalk G, Hawco SM, Prior KA (2000) Ecosystem Plan for Pelee Island. In: Proceedings of a Conference on the Biology and Management of Species and Habitats at Risk. B.C. Ministry of Environment, Lands and Parks, Victoria, B.C. and University College of the Cariboo, Kamloops, B.C, Kamloops, BC, pp 903– 908
- Fourcade Y, Engler JO, Rödder D, Secondi J (2014) Mapping Species Distributions with MAXENT Using a Geographically Biased Sample of Presence Data: A Performance Assessment of Methods for Correcting Sampling Bias. PLOS ONE 9:e97122. https://doi.org/10.1371/journal.pone.0097122
- Hirzel AH, Le Lay G (2008) Habitat suitability modelling and niche theory. J Appl Ecol 45:1372–1381. https://doi.org/10.1111/j.1365-2664.2008.01524.x
- Hossie TJ (2018) Recovery Strategy for Small-mouthed Salamander (Ambystoma texanum) and Unisexual Ambystoma Small-mouthed Salamander dependent population (Ambystoma laterale - texanum) in Ontario. Ontario Ministry of Natural Resources and Forestry, Peterborough, Ontario.
- Kimmig SE, Beninde J, Brandt M, et al (2020) Beyond the landscape: Resistance modelling infers physical and behavioural gene flow barriers to a mobile carnivore across a metropolitan area. Mol Ecol 29:466–484. https://doi.org/10.1111/mec.15345
- Kramer-Schadt S, Niedballa J, Pilgrim JD, et al (2013) The importance of correcting for sampling bias in MaxEnt species distribution models. Divers Distrib 19:1366– 1379. https://doi.org/10.1111/ddi.12096
- Larson MA, Thompson FR, Millspaugh JJ, et al (2004) Linking population viability, habitat suitability, and landscape simulation models for conservation planning. Ecol Model 180:103–118. https://doi.org/10.1016/j.ecolmodel.2003.12.054
- Lobo JM, Jiménez-Valverde A, Hortal J (2010) The uncertain nature of absences and their importance in species distribution modelling. Ecography 33:103–114
- Ministry of Natural Resources and Forestry (MNRF) (2017) Blue Racer, Lake Erie Watersnake and Small-mouthed Salamander. Ministry of Natural Resources and Forestry, Peterborough, ON
- Nature Conservancy of Canada (NCC) (2008) Management Guidelines: Pelee Island Alvars. NCC - Southwestern Ontario Region, London, ON
- Office of the Auditor General of Ontario (2020) Value-for-Money Audit: Conserving the Natural Environment with Protected Areas. Office of the Auditor General of Ontario, Toronto, ON
- Rudnick DA, Ryan SJ, Beier P, et al (2012) The role of landscape connectivity in planning and implementing conservation and restoration priorities. Ecological Society of America, Rocky Mountain Research Station, Fort Collins, CO
- Semlitsch RD, Bodie JR (2003) Biological Criteria for Buffer Zones around Wetlands and Riparian Habitats for Amphibians and Reptiles. Conserv Biol 17:1219–1228. https://doi.org/10.1046/j.1523-1739.2003.02177.x
- Semlitsch RD, Walls SC, Barichivich WJ, O'Donnell KM (2017) Extinction Debt as a Driver of Amphibian Declines: An Example with Imperiled Flatwoods Salamanders. J Herpetol 51:12–18. https://doi.org/10.1670/16-090
- Stratmann TSM, Barrett K, Floyd TM (2016) Locating Suitable Habitat for a Rare Species: Evaulation of a Species Distribution Model for Bog Turtles (Glyptemys muhlenbergii) in the Southeastern United States. Herpetol Conserv Biol 11:199– 213
- Thorn JS, Nijman V, Smith D, Nekaris K a. I (2009) Ecological niche modelling as a technique for assessing threats and setting conservation priorities for Asian slow lorises (Primates: Nycticebus). Divers Distrib 15:289–298. https://doi.org/10.1111/j.1472-4642.2008.00535.x
- Thuiller W, Münkemüller T (2010) Habitat suitability modelling. In: Effects of Climate Change on Birds. Oxford University Press Inc., New York, NY, pp 77–85
- Udyawer V, Somaweera R, Nitschke C, et al (2020) Prioritising search effort to locate previously unknown populations of endangered marine reptiles. Glob Ecol Conserv 22:e01013. https://doi.org/10.1016/j.gecco.2020.e01013
- Van Drunen SG, Linton JE, Bogart JP, et al (2020) Estimating critical habitat based on year-round movements of the endangered Jefferson Salamander (Ambystoma jeffersonianum) and their unisexual dependents. Can J Zool 98:117–126
- Ward M, Hossie TJ (2020) Do Existing Constructed Ponds on Pelee Island, Ontario Match the Habitat Requirements of Endangered Ambystoma Larvae? Wetlands 40:2097–2108. https://doi.org/10.1007/s13157-020-01364-8
- Yackulic CB, Chandler R, Zipkin EF, et al (2013) Presence-only modelling using MAXENT: when can we trust the inferences? Methods Ecol Evol 4:236–243. https://doi.org/10.1111/2041-210x.12004

Supplementary Information

Table S1: Resistance values created for *Ambystoma* salamanders on Pelee Island, ON, Canada, based on work by Compton et al. (2007). A resistance value of 1 indicates minimal resistance (i.e., most easy to travel through), and a value of 20 indicates maximum resistance (i.e., movement completely impeded). This table also outlines how we grouped land cover types in our analyses to reduce the dimensionality of this variable.

Table S2: Proportion of land cover types across Pelee Island, ON, according to the classification used in these models.

Table S3: AUC values of Maxent models for *Ambystoma texanum* and unisexual *Ambystoma*. Training data AUC is calculated from training data selected from the data set, and test data AUC is the real test of the model's predictive power.

Figure S1: Predicted habitat suitability for adult and juvenile salamanders on Pelee Island, ON, Canada, using spatially filtered localities (from 2015-2019) and environmental predictor variables, including distance to breeding site, land cover type, soil type, and elevation. Left panel: Predicted habitat suitability based on all adult *Ambystoma* salamander locations (*n* = 369). Right panel: Predicted habitat suitability based on all juvenile *Ambystoma* salamander locations (*n* = 179). Green indicates areas with predicted high habitat suitability and red indicate areas with low habitat suitability.

Figure S2: Predicted habitat suitability for three seasons on Pelee Island, ON, Canada, using spatially filtered localities (from 2015-2019) and environmental predictor variables, including distance to breeding site, land cover type, soil type, and elevation. Left panel: Predicted habitat suitability based on *Ambystoma* salamander locations from March and April 2015-2019 (*n* = 190). Center panel: Predicted habitat suitability based on *Ambystoma* salamander locations from May – July 2015-2019 (*n* = 233). Right panel: Predicted habitat suitability based on *Ambystoma* salamander locations from August and September 2015-2019 ($n = 136$). Green indicates areas with predicted high habitat suitability and red indicates areas with low habitat suitability.

Figure S3: Predicted habitat connectivity for adult and juvenile life stages on Pelee Island, ON, Canada, using spatially filtered localities (from 2015-2019) and a land cover raster grid using resistance values based on Compton et al. 2007 (see Table S1). Left panel: Connectivity based on adult *Ambystoma* salamander locations (*n* = 369). Right panel: Connectivity based on juvenile *Ambystoma* salamander locations (*n* = 179). Green indicates well-connected areas and red indicates areas with low connectivity.

Figure S4: Predicted habitat connectivity comparing seasons on Pelee Island, ON, using spatially filtered localities and a land cover raster grid using resistance values based on Compton et al. 2007 (see Table S1). Left panel: Connectivity based on *Ambystoma* salamander locations from March and April 2015-2019 (*n* = 190). Center panel: Connectivity based on *Ambystoma* salamander locations from May – July 2015-2019 (*n* = 233). Right panel: Connectivity based on *Ambystoma* salamander locations from August and September 2015-2019 (*n* = 136). Green indicates well-connected areas, with orange and red indicating areas with low connectivity.

Figure S5: Effect of distance to breeding site (m) on predicted habitat suitability for adult and juvenile life stages of *Ambystoma* salamanders on Pelee Island, ON, Canada collected from 2015- 2019. Dashed line depicts the relationship for adult *Ambystoma* salamanders. Solid line depicts the relationship for juvenile *Ambystoma* salamanders. Higher values on y-axis represent higher habitat suitability.

Figure S6: Effect of distance to breeding site (m) on predicted habitat suitability *Ambystoma* salamanders on Pelee Island, ON, Canada across seasons using data collected from 2015-2019. Dotted line indicates model response based on *Ambystoma* salamander locations from March-April (*n* = 190). Dashed line indicates model response based on *Ambystoma* salamander locations from May-July (*n* = 233). Solid line indicates model response based on *Ambystoma* salamander locations from August-September (*n* = 136). Higher values on y-axis represent higher habitat suitability.

Figure S7: Effect of tested environmental variables on predicted habitat suitability for *Ambystoma* salamanders (*n* = 476) on Pelee Island, ON, Canada collected from 2015-2019. A) Effect of land cover classifications on Maxent model. Land cover classes were 1: Wetland, 2: Forest, 3: Tallgrass Community, 4: Agricultural, 5: Transportation, 6: Human Disturbance, 7: Open Water. B) Effect of elevation on Maxent model. C) Effect of soil type on Maxent model. Soil classes were 1: Eastport Sand, 2: Farmington Loam, 3: Brookston Clay, 4: Toledo Clay, 5: Marsh. Higher values on y-axis represent higher habitat suitability.

Figure S8: Effect of land cover type on predicted habitat suitability of *Ambystoma* salamanders on Pelee Island, ON, Canada collected from 2015-2018, based on tested genomotypes. A) locations of *Ambystoma texanum* (TT) individuals (*n* = 37). B) locations of unisexual diploid (LT) individuals (*n* = 286). C) locations of unisexual triploid (LTT) individuals (*n* = 156). Land cover classes were $1 - W$ etland, $2 -$ Forest, $3 -$ Tallgrass Community, $4 -$ Agricultural, $5 -$ Transportation, $6 -$ Human Disturbance, $7 -$ Open Water. Higher values on y-axis represent higher habitat suitability. Note the different scales on the y-axis.

Figure S9: Effect of soil type on predicted habitat suitability of *Ambystoma texanum* (TT) salamanders on Pelee Island, ON, Canada collected from 2015-2018. Soil type classes were 1- Eastport Sand, 2- Farmington Loam, 3- Brookston Clay, 4- Toledo Clay, 5- Marsh. Higher values on y-axis represent higher habitat suitability.

Figure S10: Effect of land cover type on predicted habitat suitability of *Ambystoma* salamanders on Pelee Island, ON, Canada collected from 2015-2019, based on tested life stages. A) locations of adult salamanders (*n* = 369). B) locations of juvenile salamanders (*n* = 179). Land cover classes were 1: Wetland, 2: Forest, 3: Tallgrass Community, 4: Agricultural, 5: Transportation, 6: Human Disturbance, 7: Open Water. Higher values on y-axis represent higher habitat suitability.

Figure S11: Effect of land cover type on predicted habitat suitability of *Ambystoma* salamanders on Pelee Island, ON, Canada collected from 2015-2019, based on tested seasons. A) locations collected in March and April ($n = 190$). B) locations collected from May-July ($n = 233$). C) locations collected in August and September ($n = 136$). Land cover classes were 1 – Wetland, 2 – Forest, 3 – Tallgrass Community, 4 – Agricultural, 5 – Transportation, 6 – Human Disturbance, 7 – Open Water. Higher values on y-axis represent higher habitat suitability. Note the different scales on the y-axis.

Figure S12: Distribution of habitat suitability of *Ambystoma* salamanders across Pelee Island, ON, Canada collected from 2015-2019, based on predicted suitability of individual 10x10m grid cells ($n = 173,764$) from model output using all salamander location data ($n = 476$). Suitability values ranged from 0 to 1: values <0.2 represented unsuitable habitat, values between 0.2 and 0.4 represented low suitability, values between 0.4 and 0.6 represented intermediate suitability and values >0.6 represented high suitability.

Figure S13: Distribution of habitat suitability of *Ambystoma* salamanders across Pelee Island, ON, Canada collected from 2015-2018, based on predicted suitability of individual 10x10m grid cells (*n* =173,764) from model outputs of tested genomotypes. Black represents all unisexual diploid (LT) locations ($n = 286$); Dark gray represents all unisexual triploid (LTT) locations ($n =$ 156); Light gray represents all *Ambystoma texanum* (TT) locations (*n* = 37). Suitability values ranged from 0 to 1: values <0.2 represented unsuitable habitat, values between 0.2 and 0.4 represented low suitability, values between 0.4 and 0.6 represented intermediate suitability and values >0.6 represented high suitability.

Figure S14: Distribution of habitat suitability of *Ambystoma* salamanders across Pelee Island, ON, Canada collected from 2015-2019, based on predicted suitability of individual 10x10m grid cells (*n* =173,764) from model outputs of tested life stages. Black represents all adult salamander location data ($n = 369$); Gray represents all juvenile salamander location data ($n = 179$). Suitability values ranged from 0 to 1: values <0.2 represented unsuitable habitat, values between 0.2 and 0.4 represented low suitability, values between 0.4 and 0.6 represented intermediate suitability and values > 0.6 represented high suitability.

Figure S15: Distribution of habitat suitability of *Ambystoma* salamanders across Pelee Island, ON, Canada collected from 2015-2019, based on predicted suitability of individual 10x10m grid cells (*n* =173,764) from model outputs of tested seasons. Black represents all locations collected in March and April ($n = 190$); Dark gray represents all locations collected from May-July ($n =$ 233); Light gray represents all locations collected in August and September ($n = 136$). Suitability values ranged from 0 to 1: values <0.2 represented unsuitable habitat, values between 0.2 and 0.4 represented low suitability, values between 0.4 and 0.6 represented intermediate suitability and values >0.6 represented high suitability.